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Cold atmospheric pressure plasma for treatment of chronic wounds: drug or medical device?

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Corresponding Author:	Ojan Assadian, MD, DTMH Medizinische Universität Wien Vienna, AUSTRIA
Corresponding Author Secondary Information:	
Corresponding Author's Institution:	Medizinische Universität Wien
Corresponding Author's Secondary Institution:	
First Author:	Axel Kramer, MD, PhD
First Author Secondary Information:	
Order of Authors:	Axel Kramer, MD, PhD Barbara R Conway, PhD Konrad Meissner, MD, PhD Fritz Scholz, PhD B H Rauch, PhD Anke Moroder Alexander Ehlers Alfred J. Meixner, PhD Claus-Dieter Heidecke, MD, PhD Lars Ivo Partecke, MD Manfred Kietzmann, PhD Ojan Assadian, MD, DTMH
Order of Authors Secondary Information:	
Response to Reviewers:	Authors' Reply to Reviewer comments Ref.: Ms. No. jowc.2016.0161 Cold atmospheric pressure plasma for treatment of chronic wounds: drug or medical device? Journal of Wound Care I would be grateful if you could highlight the changes you have made to the text in yellow (not using tracked changes) and include a summary of the revisions you have made when you resubmit your article. Reviewer #1 C1. In Germany, the society for plasma-medicine has recently undertaken activities to implement a standard for regulation of plasma devices, the DIN SPEC 91315. It is recommended to include this in the discussion.

A1: We sincerely thank the respected reviewer for pointing out to this important standard. We have included now a brief statement highlighting the DIN SPEC 91315, and we have added the standard's reference to the reference list. The added paragraph summarises the intention of the standard as follow:

"In this respect and as a first step, the technical standard DIN SPEC 91315:2014-06 was developed, which characterizes the basic physical and technical performance parameters of CAP sources to be used for bio-medical or biological experiments and for further development to become medically applicable plasma sources."

C2. Moreover, wound therapies are in general hard to place, as for instance they affect biochemical compositions of the wounds. Hence, it might be time to review our approach to medical devices in the wound care sector in general.

A2: We completely agree with this important remark. The authors discussed about adding further thoughts about this aspect, however, we had to acknowledge that a brief highlight of this matter would be insufficient, as it may leave too many legal, medical, and regulatory aspects unexplained, and a full discussion on the difficulties with current registration procedures, particularly for wound management devices, would inflate the manuscript inappropriately. Therefore, we kindly ask the respected reviewer not to elaborate on this aspect and keep the topic for a separate manuscript in future.

Reviewer #2

C1. This manuscript has a large number of references, often with 4 or 6 supporting each statement. In reviewing titles of the references and searching for abstracts and papers, it appears that a number of these references may not be primary sources, but are perhaps reviews that have referenced other primary sources supporting the statements. Where definitive statements are made, this reviewer recommends that primary sources should be cited. An example, among others, is the section in Background covering mutagenicity, supported by references 53-56. 53 and 54 have, respectively, the following titles: "Hydroxyl radicals attack metallic gold" and "A solid state redox buffer as interface of solid-contact ISE - a strategy to improve the reproducibility and stability of potentials". These do not appear to be primary sources for the scientific data on mutagenicity. Reference 57 also does not appear, based on the abstract, to be a primary source of data on "the latter was as result of directly exposing DNA". Reference 58 does not appear, based on the abstract, to refer to coagulation but rather it refers to skin disinfection and the thermal aspects of this. Reference 59 is a report of an in vitro study on plasma, ozone and hydrogen peroxide using a device that is identified by reference to previous papers (refs 22 and 34) which does not appear to support the assertion that this is a plasma skin regeneration device. The device is not identifiable; it should be identified and supported by clinical references if they are available. Reference 60 is not the MEDDEV Borderline Guideline. Reference 61 is not a description of the decision by the European Court. The references cited stop at 61 in the references list but the numbering in the manuscript continues up to 90.

A1: We thank the respected reviewer for pointing out the many mistakes with the reference list. Indeed, we are inconsolable about this inexcusable error! After re-checking the references, we have noticed that the complete reference list was muddled up with previous versions. Regretfully, it seems that multiple versions of the manuscript and inadequate use of the automated reference manager software have resulted in chaotic references. We have completely re-arranged the references, which should be all corrected now. Again, we whole heartily apologize for the inconvenience caused.

Additional Information:

Question

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A. Kramer¹, B.R. Conway², K. Meissner³, F. Scholz⁴, B.H. Rauch⁵, A. Moroder⁶, A. Ehlers⁶, A. J. Meixner⁷, C.-D. Heidecke⁸, L.I. Partecke⁸, M. Kietzmann⁹, O. Assadian^{10*}

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A. Kramer¹, B.R. Conway², K. Meissner³, F. Scholz⁴, B.H. Rauch⁵, A. Moroder⁶, A. Ehlers⁶, A. J. Meixner⁷, C.-D. Heidecke⁸, L.I. Partecke⁸, M. Kietzmann⁹, O. Assadian^{10*}

¹Institute of Hygiene and Environmental Medicine, University Medicine Greifswald, Germany

² Department of Pharmacy, School of Applied Sciences, University of Huddersfield, United Kingdom

³ Department of Anesthesiology and Intensive Medicine, University Medicine, Greifswald, Germany

⁴ Institute of Biochemistry, Working Group Analytical Chemistry and Environmental Chemistry, University Medicine Greifswald, Germany

⁵ Department of Pharmacology, Center of Drug Absorption and Transport, University Medicine Greifswald, Germany

⁶ Ehlers, Ehlers & Partner Healthcare Law Firm Munich, Germany

⁷ Institute of Physical and Theoretical Chemistry Tübingen, Germany

⁸ Department of Surgery, University Medicine Greifswald, Germany

⁹ Department of Pharmacology, Toxicology and Pharmacy, University of Veterinary Medicine Hannover, Foundation, Hannover, Germany

¹⁰ Institute for Hospital Epidemiology and Infection Control, Medical University of Vienna, Vienna, Austria

***Corresponding author:**

Prof. Ojan Assadian, MD, DTMH

Institute for Hospital Epidemiology and Infection Control

Medical University of Vienna, Vienna General Hospital

Waehringer Guertel 18-20

1090 Vienna, Austria

E-Mail: ojan.assadian@meduniwien.ac.at

Declaration of interest

None of the authors have any competing interests.

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Abstract

Background. The use of cold atmospheric pressure plasmas (CAPs) as a new therapeutic option for chronic wounds appears to be promising. Currently, uncertainty exists regarding their classification as medical device or medical drug.

Objective. Because the classification of CAP has medical, legal, and economic consequences as well as implications for the level of pre-clinical and clinical testing, the correct clarification is not an academic debate, but an ethical need.

Method. A multidisciplinary team of researchers and lawyers has analyzed the physical and technical characteristics as well as legal conditions of the biological action of CAP.

Results. It is concluded that the mode of action of the locally generated CAP with its main active components, being different radicals, is pharmacological and not physical in nature.

Conclusion. Depending on the intended use, CAP should be classified as a drug, which is generated by use of a medical device directly at the point of therapeutic application.

Background

Highly energetic physical plasmas comprise a mixture of reactive ionized particles, which can be physically adjusted to body temperature by mixing protons, resulting in so-called “cold plasma”. Cold atmospheric pressure plasmas (CAPs) are generated under atmospheric pressure at ambient temperatures ranging from 20°C to 50°C.^{1, 2}

Interest in medical applications of CAPs is rapidly increasing. CAPs ignited a technological spark in industry, biotechnology, and in the field of hygiene, for indications where the use of antimicrobial active agents or heat sterilization is limited. This includes decontamination of thermo-labile medical devices, food, packaging materials, waste-water, or indoor air.³ For such applications, the question does not arise whether CAPs are a drug or a medical device. On the other hand, plasma has shown promising antiseptic results on skin and mucosal membranes in infection-related diseases in dermatology⁴⁻⁶ and dentistry⁷, and has also been found to exert anti-carcinogenic effects similar to those of oxygen radicals, mainly by dose-dependent anti-proliferative⁸ and apoptotic activity.⁹⁻¹³ First reports on the use of argon plasma for tumor removal date back to 1989¹⁴, followed by successful ablation of non-neoplastic Barrett's mucosa¹⁵ and other neoplastic diseases.¹⁶⁻²¹ A plasma-activated medium has even been claimed to exert anti-tumor effects.^{22, 23}

Results of in-vitro,²⁴ in-vivo²⁴⁻²⁸ and initial clinical studies on chronic wounds of domestic animals^{29, 30} and humans³¹⁻³⁵ indicate that CAP may be a promising alternative to conventional treatment options, based on the following biological premises:

- A wound cannot heal as long as it is infected³⁶. CAP may support wound healing through its antiseptic efficacy in vitro³⁷⁻³⁹, ex vivo⁴⁰, in vivo⁴¹ and in humans.^{31, 32, 42-44}
- CAP may facilitate the transformation of the chronic wound from a stagnating wound to an acute healing wound, i.e. by inflammatory^{45,46} and proliferation-supporting stimuli^{47,48}, including stimulation of neovascularization.^{29,49,50}

However, while the medical applications of CAP are promising, it is vital to ensure the safety and reliability of applied CAPs in humans before it may be considered for routine clinical

practice. Therefore, aside from the clinical effectiveness, the safety for patients and users is an important consideration during the registration and approval processes with health authorities. Based on the intended use and the potential risks associated with a medical intervention, be it supportive, diagnostic, or therapeutic, products may be classified as medical devices, or medical drugs.

In this respect and as a first step, the technical standard DIN SPEC 91315:2014-06 was developed, which characterizes the basic physical and technical performance parameters of CAP sources to be used for bio-medical or biological experiments and for further development to become medically applicable plasma sources.⁵¹ However, findings on mutagenicity⁵²⁻⁵⁴ and transient expression of pre-mutagenic active compounds, even though the latter was as result of directly exposing DNA⁵⁵, suggest that the current risk assessment for plasma applications is still incomplete and there is a need to consider also energy, penetration depth, and the body's detoxification capacity must also be considered. In addition to classification issues, both pre-clinical and clinical investigations need to be preceded by tests of clinical effectiveness and the exclusion of chronically toxic, mutagenic, and carcinogenic risks.

The final decision on the correct classification of CAP has not only medical, legal, and economic consequences, but also fundamental implications for the required level of pre-clinical testing, and future development of this new technology. To date, the final classification of CAPs as a medical drug or medical device has been the subject of academic debate, resulting in uncertainty on the required test data for registration with the respective health authorities in Europe and the US, product liability, manufacturers' responsibility, and future economic strategies. Therefore, a multidisciplinary team of physicians, surgeons, pharmacists, physicists, and lawyers familiar with the medical application of CAPs has critically evaluated the requirements for the classification of cold atmospheric pressure plasmas as a medical drugs or medical devices from different perspectives.

Differentiation between drug and medical device

In Europe and the US, medical drugs and medical devices are regulated by separate laws, which cannot be applied simultaneously. Dual labeling is generally not accepted by drug administration agencies and thus entities are regulated by either the medical devices or medicines legislation, but not both. Historically, medical drugs were distinguished from medical devices by their pharmacological, metabolic, and/or immunologic effects, while the mode of action of medical devices was predominately based on physical aspects.

Furthermore, based on the potential risk associated with the use of medical devices, they can be distinguished into 3 classes. Class I medical devices are defined as devices without risks to humans due to low invasiveness, temporary use (≤ 60 min), and application with non-critical skin contact. Class IIa medical devices bear potential risks as they are associated with moderate invasiveness, short-term uninterrupted or repeated use (≤ 30 d) within the human body (ophthalmological, intestinal, or surgically created body cavities). Class IIb medical devices have potential elevated risks, elicit systemic effects, or are intended for use beyond 30 days (i.e. non-invasive birth control). Finally, class III medical devices have the highest potential risks, e.g. technical solutions for long-term drug application, pacemakers, implants, or invasive birth control devices.

However, the intention of preventing or treating illnesses may apply to both medical drugs and medical devices, and therefore does not qualify as a distinguishing characteristic between the two. Thus, the only reason to objectively differentiate between a drug and a medical device is its pharmacologic, immunologic or metabolic effect, compared to the physical effect of the medical device, which may merely support pharmacologic, immunologic or metabolic effects. Pertaining to CAP devices, their categorization as drugs or medical devices therefore depends solely on their mode of action. For example, surgical CAP devices used for cutting or coagulating tissue in humans (so-called "beamers")⁵⁶ or plasma skin regeneration devices⁵⁷, were previously approved as medical devices class IIb, and based on their physical (thermal) mode of action.

In contrast, the MEDDEV-Borderline guideline⁵⁸ defines a pharmacologic effect as an interaction between molecules of the substance in question and a cellular component, such as a receptor, which either elicits a direct response or blocks another one in response to a third agent. The view therefore does not specifically demand an interaction of substance molecules with “cellular components of the host”, but merely requires “cellular components” only. A recent decision of the European Court⁵⁹ supports the definition of a pharmacologic effect of a substance as an interaction with any cellular components within the host’s body, including foreign targets cells like bacteria, viruses, or parasites. This opinion is also supported by the European directive 2004/27/EG, a revised version of the directive 2001/83/EG section 2, subsection 2, which elevates the MEDDEV-Borderline guideline⁵⁸ to the instrument of choice in cases of uncertainty for defining a new technology.

However, where pharmacological aspects outweigh the physical therapeutic use of CAP, its classification requires a more complex consideration. In the case of CAP, the transferability of obvious physical and biological effects between different CAP devices is complex due to various potential mixtures of reactive species and the application of energy during plasma application.

Physical and chemical characteristics of CAP

The different available technologies relevant for plasma medicine can be categorized either as direct or indirect physical plasmas. In the case of direct plasmas, the patient’s body serves as the second electrode through an electric voltage field between the head of the device and the skin surface to be treated, i.e. dielectrically impaired discharges (DBE).^{60,61} Indirect plasmas (plasma jets) arise between two electrodes, e.g., within a hand-piece, and are conveyed outward by an intense gas stream.^{1,62} Depending on the type of carrier gas (i.e., argon or helium containing atmospheric pressured ambient air plasma), the energy supply for plasma generation and the type of plasma generation, the plasma may appear as a visible flame-like beam from a nozzle.¹ In both directly and indirectly generated plasma, the plasma contains charged particles (electrons, ions), excited atoms and molecules (i.e.,

singulett-oxygen), free radicals (atoms or molecules containing an unpaired electron), photons and electromagnetic fields, leading to the emission of visible UV or VUV radiation. The chemical composition and the physical characteristics of the generated plasma significantly depend on a number of variables such as pressure, gas mixture, design of the device, physical stimuli, and others. The reactive compounds, which become biochemically active, emerge either during the generation of the plasma in due course of interaction with molecules of the surrounding air, and/or with the medium, the bodily fluid, or the tissue to be treated.⁶³⁻⁶⁵

In addition to the direct plasma effects due to the composition of the CAP at the effect site, there are also subsequent secondary actions within the tissue, based on radical formation generated by CAP, similar to physiologically generated radicals. Hydroxyl radicals ($\bullet\text{OH}$) may act as second messengers in T-cell activation,⁶⁶ and induce apoptosis.⁶⁷ Nitrogen monoxide (NO) acts as a vasodilator, influences tissue oxygen metabolism,⁶⁸ and can prevent excessive amplification of Th1 cells.⁶⁹ NO and peroxynitrite (ONOO^-) promote angiogenesis, influence cell proliferation and differentiation.^{70,71} These examples of reactive oxygen and nitrogen species are also expected in CAP or in interaction of plasma with fluids,^{72, 73} and could substantially contribute to generating the biological effects of CAP. The major difference between plasma species and endogenous radical production is the increased amounts of oxygen and nitrogen reactive species during CAP exposure. Although in endogenous metabolism, radicals are produced, increased generation of oxygen species can lead to direct cell damage.^{74,75} The effect of electrons and ions in generating radicals in CAPs are probably not necessarily of radical nature, but rather consequential reactions of the body following oxidative stress. Radicals, in particular, can exert ionizing effects on tissue components due to their tendency towards electron donation or uptake. Radical-driven reactions attack a variety of chemical bonds during this process.⁷⁶

Hydrogen peroxide can be considered a pharmacologically relevant oxidation product.⁷⁷ The reactions following plasma use described above compare to the phenomena seen after the exposition of mononuclear blood cells to culture media, in that similar anti-proliferative and cytotoxic effects could be elicited by long-term CAP treatment of cell suspensions.⁷⁸ This is in

line with demonstrable anti-microbial effects of saline solution⁷³ and water⁷⁹ following prior treatment with CAP. So far, it remains questionable whether the “induction of the phosphate starvation response regulon PhoP by Argon plasma”⁸⁰ is a primary or secondary pharmacological effect.

The enhanced oxygen radical reactivity has two causes:

(a) In a thermodynamic capacity, oxygen radicals are relatively strong oxidants.

($E_{\text{OH}/\text{H}_2\text{O}}^{\ominus} = 2.813 \text{ V}$ is significantly larger than the standard potentials of hydrogen peroxide and oxygen: $E_{\text{H}_2\text{O}_2/\text{H}_2\text{O}}^{\ominus} = 1.763 \text{ V}$, $E_{\text{O}_2/\text{H}_2\text{O}}^{\ominus} = 1.229 \text{ V}$.)

(b) In a kinetic manner, oxygen radicals are, just like many other simply charged gas radicals, quite reactive, because they contribute to one-electron reactions, which in general can occur faster than complex redox reactions with oxidation number changes of more than ± 1 . Due to the known relationship between linear-free enthalpies, high constants for the speed of oxidation correlate with high standard potentials. Superoxide radicals bear a rather low oxidation potential (around -0.3 V), and therefore are only weak oxidants and rather strong reductive agents. Nevertheless, the reactions of the superoxide ion in water are quite fast, because protonation leads to generation of the hydrogen peroxide radical HO_2 , and the standard potentials for the formation of H_2O_2 or H_2O are rather high: $E_{\text{HO}_2/\text{H}_2\text{O}_2}^{\ominus} = 1.44 \text{ V}$, $E_{\text{HO}_2/\text{H}_2\text{O}}^{\ominus} = 1.65 \text{ V}$. The risk potential of OH radicals is furthermore emphasized by the fact that they even attack seemingly inert substances, such as gold, liberating gold ions, rather than forming a stable surface of gold oxide, which could be expected following normal thermodynamic rules.^{81, 82}

Classification of CAPs as medical drug or medical device

The classification of CAPs depends on its intended use. It is crucial to determine whether or not solely physically (thermal, such as in coagulation), or pharmacologically active substrates are being generated as part of the main effect. Even in the latter case, however, CAPs can

still be classified as medical device when used for cleaning of artificial surfaces, e.g. biofilm removal on dental implants.

All therapeutic effects of CAP reported to date are mainly based on effects of radical-derived reactive products,^{73,74,83,84} which are associated with a pharmacological/immunological effect. Physical effects such as increased body temperature, electromagnetic fields, and (CAP source-dependent) UV or VUV radiation can potentially enhance these effects. However, therapeutic effects in healing of chronic wounds⁸⁵ cannot be explained by physical effects alone. Moreover, the clinical course of chronic wound healing after the application of plasma in domestic animals and humans suggests that the plasma effect may trigger an intermediate phase of acute inflammation.⁸⁶ The underlying immunological and biological mechanisms are not yet fully understood, but induction of integrins,⁸⁷ NO,⁸⁸ and increased phosphorylation of β -Catenin⁸⁹ may indicate pharmacological and immunological effects of CAPs.

However, the complex composition of CAP and its generation within the plasma source at the time of application further corroborates its systematic classification within the classic drug category. In comparison with “classic” medical drugs, this may have the following consequences:

- The CAP compounds cannot be prepared and stored in a carrier substrate in defined doses with detailed knowledge about the exact composition before application on a patient.
- For CAP, storage times cannot be defined, as required for drugs or certain medical devices such as surgical or medical examination gloves. Instead, routine technical service and maintenance of the plasma-generating source will be required.
- Newly generated plasma intended for medical use can be uniquely different from **another plasma** in terms of its composition, depending on the type of plasma generation, the carrier gas used and environmental conditions. Therefore, the actual plasma for each current use needs to be thoroughly described in terms of its active components. During each use, narrow margins of tolerance need to be defined and

followed. Quantifiable parameters of action are: energy application (J/m^2), temperature, field strength, UV spectrum, gas uses, and, associated with these, radicals in plasma. However, due to technical limitations, accurate quantification of radicals is not currently feasible. Moreover, it is technically challenging to maintain the energy application constant for a particular plasma source.

- Similar to a drug, CAPs intended for therapeutic application need to be composed of a defined content, which is unaltered by changes in environmental conditions, such as humidity of the ambient air. Therefore, plasma application needs to be independent of the surrounding atmosphere.

Conclusion

CAP as a new therapeutic option for chronic wounds or for neoplastic formations appears to be promising. Nevertheless, potentially deleterious long-term effects need to be ruled out. This applies particularly to mutagenic and carcinogenic risks of long-term plasma application. Therefore, before CAPs can safely be used in clinical applications for wound and skin treatment or treatment of tumors, it is imperative to further evaluate the interaction of living pro- and eukaryotic cells with CAP.

Analysis of physical and pharmacological properties of CAP supports its classification as a drug, which is generated by use of a medical device directly at the point of therapeutic application. In contrast to conventional drugs, CAP and its biologically active compounds are generated at the time of application by use of a medical device. Therefore, the effective plasma itself, which is generated at the treatment site could be regarded as a local redox modulator, which stimulates the cells and thus represents an applied pharmacological principle. The mode of action of the locally generated CAP with its main active components – the different radicals – is pharmacological and not physical in nature.

References

1. Weltmann, K.D., Kindel, E., Woedtke, T. von, Hähnel, M., Stieber, M., Brandenburg, R. **Atmospheric-pressure plasma sources: Prospective tools for plasma medicine.** Pure Appl Chem 2010; 82: 1223–37.
2. Isbary, G., Shimizu, T., Li, Y.F., Stolz, W., Thomas, H.M., Morfill, G.E., Zimmermann, J.L. **Cold atmospheric plasma devices for medical issues.** Expert Rev Med Dev 2013; 10: 367–77.
3. Kramer, A., Bekeschus, S., Matthes, R., Bender, C., Stope, M.B., Napp, M., et al. **Cold physical plasmas in the field of hygiene—relevance, significance, and future applications.** Plasma Proc Polym 2015; 12: 1410–22.
4. Emmert, S., Brehmer, F., Hänßle, H., Helmke, A., Mertens, N., Ahmed R., et al. **Atmospheric pressure plasma in dermatology: Ulcus treatment and much more.** Clin Plasma Med 2013; 1 (1): 24–9.
5. Klebes, M., Lademann, J., Philipp, S., Ulrich, C., Patzelt, A., Ulmer, M., et al. **Effects of tissue-tolerable plasma on psoriasis vulgaris treatment compared to conventional local treatment: A pilot study.** Clin Plasma Med 2014; 2 (1): 22–7.
6. Isbary, G., Shimizu, T., Zimmermann, J.L., Heinlin, J., Al-Zaabi, S., Rechfeld, M., et al. **Randomized placebo-controlled clinical trial showed cold atmospheric argon plasma relieved acute pain and accelerated healing in herpes zoster.** Clin Plasma Med 2014; 2 (2): 50–5.
7. Cha, S., Park, Y.S. **Plasma in dentistry.** Clin Plasma Med 2014; 2 (1): 4–10.
8. Lupu, A., Georgescu, N. **Cell death in V79-4 and He-La cell lines, induced by chemically activated cold atmospheric plasma jets.** Rom Rep Phys 2013;65: 219–29.
9. Zucker, S.N., Zirnheld, J., Bagati, A., et al. **Preferential induction of apoptotic cell death in melanoma cells as compared with normal keratinocytes using a non-thermal plasma torch.** Cancer Biol Ther 2012; 13: 1299–306.

10. Fridman, G., Shereshevsky, A., Jost ,M.M., Brooks, A.D., Fridman, A., Gutsol, A., et al. **Floating electrode dielectric barrier discharge plasma in air promoting apoptotic behavior in melanoma skin cancer cell lines.** Plasma Chem Plasma 2007; 27163–76.
11. Ahn, H.J., Kim, K.I., Kim, G., Moon, E., Yang, S.S., Lee, J.S. **Atmospheric-pressure plasma jet induces apoptosis involving mitochondria via generation of free radicals.** Plos One 2011; 6: e28154.
12. Kim CH, Bahn JH, Lee SH, Kim GY, Jun SI, Lee K, et al. **Induction of cell growth arrest by atmospheric non-thermal plasma in colorectal cancer cells.** J Biotechnol 2010; 150: 530–8.
13. Partecke LI, Evert K, Haugk J, Doering F, Normann L, Diedrich S, et al. **Tissue Tolerable Plasma (TTP) induce apoptosis in the human pancreatic cancer cell line Colo-357 in vitro and in vivo.** BMC Cancer 2012; 12: 473.
14. Brekhov, E.I. **Experimental and clinical studies and prospects of using plasma flows.** Khirurgiia (Mosk) 1989; 7: 94–6.
15. Manner H, May A, Miehlke S, Dertinger S, Wigglinghaus B, Schimming W, et al. **Ablation of non-neoplastic Barrett's mucosa using argon coagulation with concomitant esomeprazole therapy (APBANEX): a prospective multicenter evaluation.** Am J Gastroenterol 2006; 101: 1762-1769.
16. Walk RM, Snyder JA , Srinivasan P, Kirsch J, Diaz SO, Blanco FC, et al. **Cold atmospheric plasma for the ablative treatment of neuroblastoma.** J Pediatr Surg 2013; 48: 67–73.
17. Keidar M, Shashurin A, Volotskova O, Stepp MA, Srinivasan P, Sandler A, et al. **Cold atmospheric plasma in cancer therapy.** Phys Plasmas 2013; 20: 57101.
18. Köritzer J, Boxhammer V, Schäfer A, Shimizu T, Klämpfl TG, Li YF, et al. **Restoration of sensitivity in chemo-resistant glioma cells by cold atmospheric plasma.** PLoS One. 2013; 8: e64498.
19. Siu, A., Volotskova, O., Cheng, X., Khalsa, S.S., Bian, K., Murad, F., et al. **Differential effects of cold atmospheric plasma in the treatment of malignant glioma.** PLoS ONE 2015; 10: e0126313.

20. Welz, C., Emmert, S., Canis, M., Becker, S., Baumeister, P., Shimizu, T., et al. **Cold atmospheric plasma: A promising complementary therapy for squamous head and neck cancer.** PLoS ONE 2015; 10: e0141827.
21. Gay-Mimbrera, J., García, M.C., Isla-Tejera, B., Rodero-Serrano, A., García-Nieto, A.V., Ruano, J. **Clinical and biological principles of cold atmospheric plasma application in skin cancer.** Adv Ther 2016; 33: 894.
22. Kajiyama H, Utsumi F, Nakamura K, Tanaka H, Mizuno M, Toyokuni S, et al. **Possible therapeutic option of aqueous plasma for refractory ovarian cancer.** Clin Plasma Med 2016; 4: 14–8.
23. Vandamme M, Robert E, Lerondel S, Sarron V, Ries D, Dozias S, et al. **ROS implication in a new antitumor strategy based on non-thermal plasma.** Int J Cancer 2012; 130: 2185–94.
24. Kramer A, Lademann J, Bender CP, Sckell A, Hartmann B, Münch S, et al. **Suitability of Tissue Tolerable Plasmas (TTP) for the management of chronic wounds.** Clin Plasma Med 2013; 1: 11–8.
25. Nastuta, A.V., Topala, I., Grigoras, C., Pohoata, V., Popa, G. **Stimulation of wound healing by helium atmospheric pressure plasma treatment.** J Phys D: Appl Phys 2011; 44: 105204.
26. García-Alcantara, E., Lopez-Callejas, R., Morales-Ramirez, P.R., Pena-Eguiluz, R., Fajardo-Munoz, R., Mercado-Cabrera, A., et al. **Accelerated mice skin acute wound healing in vivo by combined treatment of argon and helium plasma needle.** Arch Med Res 2013; 44: 169–77.
27. Shokri, A., Khani, M.R., Bigdeli, M., Shokri, B. **Investigating effects of atmospheric-pressure plasma on the process of wound healing.** Biointerphases 2015; 10: 029504.
28. Salehi, S., Shokri, A., Khani, M.R., Bigdeli, M., Shokri, B. **Investigating effects of atmospheric-pressure plasma on the process of wound healing.** Biointerphases 2015; 10:029504.

29. Fathollah, S., Mirpour, S., Mansouri, P., Dehpour, A.R., Ghoranneviss, M., Rahimi, N., et al. **Investigation on the effects of the atmospheric pressure plasma on wound healing in diabetic rats.** *Sci Rep* 2016; 6: 19144.
30. Hung, Y.W., Lee, L.T., Peng, Y.C., Chang, C.T., Wonge, Y.K., Tung, K.C. **Effect of a nonthermal-atmospheric pressure plasma jet on wound healing: An animal study.** *J Chin Med Assoc* 2016; 79: 320–8.
31. Isbary, G., Morfill, G., Schmidt, H.U., Georgi, M., Ramrath, K., Heinlin, J., et al. **Successful and safe use of 2 min cold atmospheric argon plasma in chronic wounds: results of a randomized controlled trial.** *Br J Dermatol* 2012; 167: 404–10.
32. Ulrich, C., Kluschke, F., Patzelt, A., Vandersee, S., Czaika, V., Richter, H. et al. **Clinical use of cold atmospheric pressure argon plasma in chronic leg ulcers: a pilot study.** *J Wound Care* 2015; 24:196-203.
33. Bender, C., Hübner, N.O., Weltmann, K.D., Scharf, C., Kramer, A. **Tissue tolerable plasma and polyhexanide: Are synergistic effects possible to promote healing of chronic wounds? In vivo and in vitro results.** In: Machala Z, Hendsel K, Akishey Y, editors. *Plasma for Bio-Decontamination, Medicine and Food Security.* Dordrecht: Springer, 2012: 321-34.
34. Heinlin, J., Zimmermann, J.L., Zeman, F., Bunk, W., Isbary, G., Landthaler, M., et al. **Randomized placebo-controlled pilot study using cold atmospheric argon plasma on skin graft donor sites.** *Wound Repair Regen* 2013; 21: 800-7.
35. Isbary, G., Stolz, W., Shimizu, T., Monetti, R., Bunk, W., Schmidt, H.U., et al. **Cold atmospheric argon plasma treatment may accelerate wound healing in chronic wounds: Results of a retrospective in vivo randomized controlled study.** *Clin Plasma Med* 2013; 1:25-30.
36. Kramer, A., Assadian, O., Below, H., Willy, C. **Wound antiseptics today - an overview.** In: Willy C, editor. *Antiseptics in surgery – update 2013.* Berlin: Lindqvist, 2013: 85-111.
37. Daeschlein, G., von Woedtke, T., Kindel, E., Brandenburg, R., Weltmann, K.D., Jünger, M. **Antibacterial activity of an atmospheric pressure plasma jet against relevant**

- wound pathogens in vitro on a simulated wound environment.** Plasma Proc Polymers 2010; 7: 224–30.
38. Fricke, K., Koban, I., Tresp, H., Jablonowski, L., Schröder, K., Kramer, A., et al. **Atmospheric pressure plasma: a high-performance tool for the efficient removal of biofilms.** PloS one 2012; 7: 8,e42539.
39. Matthes, R., Bender, C., Schlüter, R., Koban, I., Bussiahn, R., Reuter, S., et al. **Antimicrobial efficacy of two surface barrier discharges with air plasma against in vitro biofilms.** PloS ONE 2013; 7(8): e42539.
40. Boekema, B., Vlig, M., Guijt, D., Hijnen, K.K., Hofmann, S.S., Smits, P.P., et al. **A new flexible DBD device for treating infected wounds: in vitro and ex vivo evaluation and comparison with a RF argon plasma jet.** J Phys D: Appl Phys 2016; 49: 044001.
41. Ermolaeva, S.A., Varfolomeev, A.F., Chernukha, M.Y., Yurov, D.S., Vasiliev, M.M., Kaminskaya, A.A., et al. **Bactericidal effects of non-thermal argon plasma in vitro, in biofilms and in the animal model of infected wounds.** J Med Microbiol 2011; 60: 75-83.
42. Isbary, G., Morfill, G.E., Schmidt, H.U., Georgi, M., Ramrath, K., Heinlin, J., et al. **A first prospective randomized controlled trial to decrease bacterial load using cold atmospheric argon plasma on chronic wounds in patients.** Brit J Dermatol 2010; 163: 78–82.
43. Isbary, G., Shimizu, T., Zimmermann, J., Hubertus, T., Morfill, G., Stolz, W. **Cold atmospheric plasma for local infection control and subsequent pain reduction in a patient with chronic post-operative ear infection.** New Microbes New Infect 2013; 1: 41–3.
44. Brehmer, F., Haenssle, H.A., Daeschlein, G., Ahmed, R., Pfeiffer, S., Görlitz, A., et al. **Alleviation of chronic venous leg ulcers with a hand-held dielectric barrier discharge plasma generator (PlasmaDerm® VU-2010): Results of a monocentric, two-armed, open, prospective, randomized and controlled trial (NCT01415622).** J Eur Acad Dermatol Venereol 2015; 29: 148–55.

45. Bender, C., Matthes, R., Kindel, E., Kramer, A., Lademann, J., Weltmann, K. D. et al. **The irritation potential of nonthermal atmospheric pressure plasma in the HET-CAM.** Plasma Proc Polym 2010; 7: 318–326.
46. Bender, C., Partecke, L.I., Kindel, E., Kramer, A., Lademann, J., Weltmann, K.D., et al. **The modified HET-CAM as a model for the assessment of the inflammatory response to tissue tolerable plasma.** Toxicol In Vitro 2011; 25: 530–7.
47. Arndt, S., Unger, P., Wacker, E., Shimizu, T., Heinlin, J., Li, Y.F., et al. **Cold atmospheric plasma (CAP) changes gene expression of key molecules of the wound healing machinery and improves wound healing in vitro and in vivo.** PLoS ONE 2013; 8: e79325.
48. Arndt, S., Landthaler, M., Zimmermann, J.L., Unger, P., Wacker, E., Shimizu, T., et al. **Effects of cold atmospheric plasma (CAP) on β -defensins, inflammatory cytokines, and apoptosis-related molecules in keratinocytes in vitro and in vivo.** PLoS ONE 2015; 10: e0120041.
49. Kisch, T., Helmke, A., Schleusser, S., Song, J., Liodaki, E., Stang, F.H., et al. **Improvement of cutaneous microcirculation by cold atmospheric plasma (CAP): Results of a controlled, prospective cohort study.** Microvasc Res 2016; 104: 55–62.
50. Kisch, T., Schleusser, S., Helmke, A., Mauss, K.L., Wenzel, E.T., Hasemann, B., et al. **The repetitive use of non-thermal dielectric barrier discharge plasma boosts cutaneous microcirculatory effects.** Microvasc Res 2016; 106: 8–13.
51. German Standardisation Committee. DIN SPEC 91315, **General Requirements For Plasma Sources In Medicine.** 2014; 1–16.
52. Kalghatgi SU, Fridman G, Cooper M, Nagaraj G, Peddinghaus M, Balasubramanian M, et al. **Mechanism of blood coagulation by non-thermal atmospheric pressure dielectric barrier discharge plasma.** IEEE Transact Plasma Sci 2007; 35:1559–66.
53. Ptasinska S, Bahnev B, Stypczynska A, Bowden M, Mason NJ, Braithwaite NSJ. **DNA strand scission induced by a non-thermal atmospheric pressure plasma jet.** Phys Chem Chem Phys 2010; 12: 7779–81.

54. Garcia-Alcantara E, López-Callejas R, Serment-Guerrero J, Peña-Eguiluz R, Muñoz-Castro AE, Rodríguez-Méndez BG, et al. **Toxicity and genotoxicity in HELA and E. coli cells caused by a helium plasma needle.** Appl Phys Res 2013; 5: 21–8.
55. Morales-Ramírez P, Cruz-Vallejo V, Peña-Eguiluz R, López-Callejas R, Rodríguez-Méndez B G, Valencia-Alvarado R, et al. **Assessing cellular DNA damage from a helium plasma needle.** Radiat Res 2013; 179: 669–73.
56. Bogle, M.A., Arndt, K.A., Dover, J.S. **Evaluation of plasma skin regeneration technology in low-energy full-facial rejuvenation.** Arch Dermatol. 2007; 143: 168–74.
57. Kilmer, S., Semchyshyn, N., Shah, G., Fitzpatrick, R. **A pilot study on the use of a plasma skin regeneration device (Portrait (R) PSR3) in full facial rejuvenation procedures.** Lasers Med Sci. 2007; 22:101–9.
58. MEDDEV 2.1/3 Rev. 3. **Guidelines relating to the application of The Council Directive 90/385/EEC on active implantable medical devices the Council Directive 93/42/EEC on medical devices.** September 2007.
59. Judgment of the Court (Fourth Chamber) of 10 July 2014. **Criminal proceedings against Markus D (C-358/13) and G. (C-181/14) References for a preliminary ruling.** Bundesgerichtshof Germany Medicinal products for human use - Directive 2001/83/EC - Scope - Interpretation of the concept of 'medicinal product' - Scope of the criterion based on the capacity to modify physiological functions - Herb and cannabinoid-based products - Not included. Joined cases C-358/13 and C-181/14.
60. Liu, C., Cui, N., Brown, N.M.D., Meenan, B.J. **Effects of DBD plasma operating parameters on the polymer surface modification.** Surf Coat Technol. 2004; 185: 311–20.
61. Bussiahn, R., Brandenburg, R., Gerling, T., Kindel, E., Lange, H., Lembke, N., et al. **The hairline plasma: An intermittent negative dc-corona discharge at atmospheric pressure for plasma medical applications.** Appl Phys Lett 2010; 96: 143701.
62. Ionin, A.A., Kochetov, I.V., Napartovich, A.P., Yuryshev, N.N. **Physics and engineering of singlet delta oxygen production in low-temperature plasma.** J Phys D Appl Phys 2007; 40: R25.

63. Kong, M.G., Kroesen, G., Morfill, G., Nosenko, T., Shimizu, T., van Dijk, J., et al. **Plasma medicine: an introductory review.** N J Phys 2009; 11: 1150.
64. Kalghatgi, S., Kroesen, G., Morfill, G., Nosenko, T., Shimizu, T., van Dijk, J., et al. **Effects of non-thermal plasma on mammalian cells.** PLoS ONE 2011; 6: e16270.
65. Tatla, S., Woodhead, V., Foreman, J.C., Chain, B.M. **The role of reactive oxygen species in triggering proliferation and IL-2 secretion in T cells.** Free Rad Biol Med 1999; 26: 14–24.
66. Ren, J.G., Xia, H.L., Tian, Y.M., Just, T., Cai, G.P., Dai, Y.R. **Expression of telomerase inhibits hydroxyl radical-induced apoptosis in normal telomerase negative human lung fibroblasts.** FEBS Letters 2001; 488, 133–8.
67. Tarricone, E., Brun, P., Vono, M., Cardin, R., Zuin, M., Martines, E., et al. **Investigation of the effects of atmospheric pressure cold plasma on human cells and tissues.** Ital J Anat Embryol 2012; 117 (suppl 2):186.
68. Fang, F.C. **Antimicrobial reactive oxygen and nitrogen species: concepts and controversies.** Nat Rev Microbiol. 2004; 2: 820–32.
69. Huang, F.P., Niedbala, W., Wei, X.Q., Xu, D., Feng, G.J., Robinson, J.H., et al. **Nitric oxide regulates Th1 cell development through the inhibition of IL-12 synthesis by macrophages.** Eur J Immunol. 1998; 28: 4062–70.
70. Hicks, K.K., Shin, J.T., Opalenik, S.R., Thompson, J.A. **Molecular mechanisms of angiogenesis: experimental models define cellular trafficking of FGF-1.** Puerto Rico Health Sci J 1996; 15:179–86.
71. Luo, J.D., Chen, A.F. **Nitric oxide: a newly discovered function on wound healing.** Acta Pharmacol Sin 2005; 26: 259–64.
72. Gaunt, L.F., Beggs, C.B., Georghiou, G.E. **Bactericidal action of the reactive species produced by gas-discharge nonthermal plasma at atmospheric pressure: A review.** IEEE Transact Plasma Sci 2006; 34: 1257–69.
73. Oehmigen, K., Winter, J., Hähnel, M., Wilke, C., Brandenburg, R., Weltmann, K.D., et al. **Estimation of possible mechanisms of Escherichia coli inactivation by plasma treated sodium chloride solution.** Plasma Proc Polym 2011; 8: 904–13.

74. Pacher, P., Beckman, J.S., Liaudet, L. **Nitric oxide and peroxynitrite in health and disease.** *Physiol Rev* 2007; 87: 315–424.
75. Halliwell, B. **Biochemistry of oxidative stress.** *Biochem Soc Transact* 2007; 35: 1147–50.
76. Dobrynin, D., Fridman, G., Friedman, G., Fridman, A. **Physical and biological mechanisms of direct plasma interaction with living tissue.** *N J Phy.* 2009; 11: 115020.
77. Bekeschus, S., Kolata, J., Winterbourn, C., Kramer, A., Turner, R., Weltmann, K.D., et al. **Hydrogen peroxide: A central player in physical plasma-induced oxidative stress in human blood cells.** *Free Radical Res* 2014; 48:542–9.
78. Bekeschus, S., Masur, K., Kolata, J., Wende, K., Schmidt, A., Bundscherer, L., et al. **Human Mononuclear Cell Survival and Proliferation is Modulated by Cold Atmospheric Plasma Jet.** *Plasma Proc Polym* 2013; 10: 706–13.
79. Naitali, M., Kamgang-Youbi, G., Herry, J., Bellon-Fontaine, M., Brisset, J. **Combined Effects of Long-Living Chemical Species during Microbial Inactivation Using Atmospheric Plasma-Treated Water.** *Appl Environm Microbiol* 2010; 76: 7662–4.
80. Winter, T., Bernhardt, J., Winter, J., Mäder, U., Schlüter, R., Weltmann, K.D., et al. **Common versus noble Bacillus subtilis differentially responds to air and argon gas plasma.** *Proteomics* 2013; 13: 2608–21.
81. Nowicka AM, Hasse U, Hermes M, Scholz F. **Hydroxyl radicals attack metallic gold.** *Ang Chem Int. Ed.* 2010; **49**: 1061–3.
82. Nowicka, A.M., Hasse, U., Sievers, G., Donten, M., Stojek, Z., Fletcher, S., et al. **A solid-state redox buffer as interface of solid-contact ISE – a strategy to improve the reproducibility and stability of potentials.** *Ang Chem Int Ed* 2010; 49: 3006–9.
83. Schmidt, A., Wende, K., Bekeschus, S., Bundscherer, L., Barton, A., Ottmüller, K., et al. **Non-thermal plasma treatment is associated with changes in transcriptome of human epithelial skin cells.** *Free Rad Res* 2013; 47: 577–92.

84. Yousfi, M., Merbahi, N., Pathak, A., Eichwald, O. **Low-temperature plasmas at atmospheric pressure: toward new pharmaceutical treatments in medicine.** *Fundam Clin Pharmacol* 2014; 28: 123–35.
85. Lademann, J., Richter, H., Schanzer, S., Patzelt, A., Thiede, G., Kramer, A., et al. **Comparison of the antiseptic efficacy of tissue-tolerable plasma and an octenidine hydrochloride-based wound antiseptic on human skin.** *Skin Pharmacol Physiol* 2012; 25:100-6.
86. Lademann, O., Richter, H., Schanzer, S., Patzelt, A., Thiede, G., Kramer, A., et al. **Application of a plasma-jet for skin antiseptis: analysis of the thermal action of the plasma by laser scanning microscopy.** *Laser Phys Lett* 2010; 7: 458–62.
87. Haertel, B., Straßenburg, S., Oehmigen, K., Wende, K., von Woedtke, T., Lindequist, U. **Differential influence of components resulting from atmospheric pressure plasma on integrin expression of human HaCaT keratinocytes.** *Biomed Res Int* 2013; 76: 761451.
88. Liebmann, J., Scherer, J., Bibinov, N., Rajasekaran, P., Kovacs, R., Gesche, R., et al. **Biological effects of nitric oxide generated by an atmospheric pressure gas-plasma on human skin cells.** *Nitric Oxide* 2011; 24: 8–16.
89. Kim, C.H., Kwon, S., Bahn, J.H., Lee, K., Jun, S.I., Rack, P.D. **Induction of cell growth arrest by atmospheric non-thermal plasma in colorectal cancer cells.** *J Biotechnol* 2010; 150: 530–8.