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# 1 **The use of bile salt micelles for the prediction of human intestinal** 2 **absorption**

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## 9 **Abstract**

10 Human intestinal absorption (HIA) will dictate biopharmaceutical performance through its  
11 influence on ADME (Absorption, Distribution, Metabolism and Elimination) and can vary  
12 significantly depending upon the nature of the compound under consideration. In this study,  
13 an *in vitro* assay method is proposed for the prediction of HIA through the measurement of  
14 drug solubility in an aqueous phase containing micellar bile salt, namely sodium  
15 deoxycholate (NaDC). A series of twenty compounds, displaying a range of physicochemical  
16 properties and known HIA values, were analysed using UV spectroscopy to determine a  
17 solubilisation ratio (SR) for each compound. A micelle/water partition coefficient ( $K_{xm/a}$ ) was  
18 calculated and then used to develop an equation through simple linear regression;  $\text{logit HIA} =$   
19  $-0.919 + 0.4618 \log K_{xm/a}$  ( $R^2 = 0.85$ ). From this equation a value for % HIA was determined  
20 which compared well with literature. Furthermore, four additional drugs were then analysed  
21 using the developed equation and found to match well with literature, confirming the  
22 suitability of the method. Using a simple, economic and robust UV bile salt assay allows  
23 prediction of human intestinal absorption and avoids many of the disadvantages of other  
24 techniques, such as animal based methods.

## 25 **Keywords**

26 Human Intestinal absorption; HIA; solubilisation; UV; bile salts

## 27 **Introduction**

28 Human intestinal absorption (HIA) is the mechanism through which drugs traverse  
29 from the intestine into the bloodstream. The vast majority of active pharmaceutical  
30 ingredients are administered orally thus it is essential that they are absorbed within the  
31 intestine to reach the intended site of action. Although it is possible to measure the percent  
32 HIA (% HIA) during clinical studies, it is far more useful to be able to predict the value much  
33 earlier on during drug development. It is for this reason that a significant amount of research  
34 has been undertaken in an attempt to develop a reliable, robust and accurate method to predict  
35 % HIA.

36 Several different predictive approaches have been undertaken, including  
37 computational (*in silico*) methods<sup>1</sup>, such as quantitative structure-activity relationships  
38 (QSARs)<sup>2</sup> and physiologically-based pharmacokinetic (PBPK) modelling<sup>3</sup>. These techniques  
39 have a clear advantage in that they remove the need for costly laboratory based experimental  
40 measurement yet their predictive ability can be limited.

41 *In vitro* models for the prediction of absorption include the application of dissolution  
42 analysis<sup>4</sup>, chromatographic analysis<sup>5</sup> and dynamic gastric models<sup>6</sup>. Many of these *in vitro*  
43 models have included the presence of physiologically relevant solvent compositions, mainly  
44 because it is known that solvent composition dictates intestinal drug solubility which, in turn,  
45 is an important factor in determining the rate, and extent, of absorption<sup>7</sup>. The specific  
46 components within human intestinal fluids that dramatically alter drug solubility are bile  
47 salts. The main biological function of bile salts is to solubilise lipids and vitamins in the  
48 intestine with a similar effect encountered for orally administered drugs. For a full review of  
49 the absorption-enhancing effects of bile salts see Ref 8<sup>8</sup>.

50 In humans, the composition of bile salts is rather complex and for the purposes of this  
51 study was simplified to consider one bile salt in particular, namely sodium deoxycholate  
52 (NaDC). NaDC is a well-characterised amphiphilic molecule which can undergo micellar  
53 aggregation<sup>9</sup>, stabilised by polar interactions<sup>10</sup>, with comparatively small aggregation  
54 numbers as a result of the rigid molecular structure<sup>11</sup>. Previous research within our group has  
55 shown that NaDC, when in the presence of drugs, will exhibit modified physicochemical  
56 properties, for example a variable (drug-specific) reduction in critical micellar concentration  
57 (CMC)<sup>12</sup>.

58 When quantifying (or comparing) enhancement in solubility for a specific drug, or  
59 series of drugs, it is possible to evaluate the solubilisation ratio (SR), where SR is equal to the  
60 moles of drug solubilised per mole of bile salt. One study in particular calculated SR for a  
61 series of steroids and then used this data to calculate micelle/water partition coefficients  
62 ( $K_{m/w}$ ) which were then correlated with octanol/water partition coefficients ( $P_{o/w}$ )<sup>13</sup>. Using  
63 this same theory as a basis for drug-NaDC measurement, this paper describes the evolution of  
64 measuring SR and then using these values as the basis to form an equation to permit  
65 prediction of % HIA, thereby presenting an *in vitro* method to predict *in vivo* behaviour.

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## 67 **Materials and Methods**

### 68 **Materials**

69 Aqueous solutions of sodium deoxycholate (NaDC), used as purchased from Sigma  
70 Aldrich, Dorset, UK (97 %), were prepared by dilution from a 20 mM stock solution with  
71 distilled water as necessary to achieve concentrations of 7, 9, 11, 13, 17 and 20 mM (i.e.  
72 always at concentrations greater than the stable micelle CMC concentration of NaDC<sup>9b</sup>). The

73 24 compounds considered in this work were: acetaminophen (99 %, Sigma Aldrich, Dorset,  
74 UK), acetyl salicylic acid (99 %, Acros organics, Geel, Belgium), alprenolol (98 %, Sigma  
75 Aldrich, Dorset, UK), amitriptyline (98 %, Sigma Aldrich, Dorset, UK), carbamazepine (99  
76 %, Sigma Aldrich, Dorset, UK), cimetidine (Sigma Aldrich, Dorset, UK), diclofenac (98 %,   
77 TCI, Europe), diphenhydramine (98 %, TCI, Europe), fenoprofen (97 %, Fluka, Dorset, UK),  
78 fluconazole (98 %, Sigma Aldrich, Dorset, UK), flurbiprofen (98 %, TCI, Europe),  
79 gemfibrozil (98 %, TCI, Europe), ibuprofen (98 %, BASF, Cheshire, UK), indomethacin (99  
80 %, Sigma Aldrich, Dorset, UK), ketoprofen (98 %, Sigma Aldrich, Dorset, UK), lidocaine  
81 (98 %, Sigma Aldrich, Dorset, UK), mannitol (98 %, Sigma Aldrich, Dorset, UK),  
82 meloxicam (98 %, TCI, Europe), naproxen (98 %, Sigma Aldrich, Dorset, UK),  
83 phenylbutazone (99 %, Sigma Aldrich, Dorset, UK), piroxicam (98 %, Sigma Aldrich,  
84 Dorset, UK), propranolol (99 %, Sigma Aldrich, Dorset, UK), quinine (96 %, Fluka, Dorset,  
85 UK), terbutaline (96 % Sigma Aldrich, Dorset, UK), used as purchased. All experimental  
86 work was conducted without altering the pH or ionic strength to avoid the formation of a  
87 surfactant-gel hydropolymer.

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## 89 **Method**

90 A calibration plot was established at each of the 6 bile salt concentrations using the  
91 Agilent Cary 60 UV-Vis spectrophotometer set at wavelength of maximum absorbance for  
92 each drug as follows (acetaminophen  $\lambda_{\max}$ . 243 nm, acetyl salicylic acid  $\lambda_{\max}$ . 295 nm,  
93 alprenolol  $\lambda_{\max}$ . 270 nm, amitriptyline  $\lambda_{\max}$ . 240 nm, carbamazepine  $\lambda_{\max}$ . 284 nm, cimetidine  
94  $\lambda_{\max}$ . 218 nm, diclofenac  $\lambda_{\max}$ . 276 nm, diphenhydramine  $\lambda_{\max}$ . 221 nm, fenoprofen  $\lambda_{\max}$ . 271  
95 nm, fluconazole  $\lambda_{\max}$ . 260 nm, flurbiprofen  $\lambda_{\max}$ . 247 nm, gemfibrozil  $\lambda_{\max}$ . 274 nm, ibuprofen  
96  $\lambda_{\max}$ . 272 nm, indomethacin  $\lambda_{\max}$ . 320 nm, ketoprofen  $\lambda_{\max}$ . 261 nm, lidocaine  $\lambda_{\max}$ . 262 nm,  
97 mannitol  $\lambda_{\max}$ . 295 nm, meloxicam  $\lambda_{\max}$ . 362 nm, naproxen  $\lambda_{\max}$ . 230 nm, phenylbutazone

98  $\lambda_{\text{max}}$ . 264 nm, piroxicam  $\lambda_{\text{max}}$ . 355 nm, propranolol  $\lambda_{\text{max}}$ . 292 nm, quinine  $\lambda_{\text{max}}$ . 332 nm,  
99 terbutaline  $\lambda_{\text{max}}$ . 280 nm), also the sample cell was thermostated at 37 °C. Separately, an  
100 excess of drug was added to 1 mL of each bile salt concentration in a microcentrifuge tube  
101 and placed in a shaking water bath for 48 hours at 37 °C, then centrifuged at 13,000 rpm,  
102 filtered and diluted using the corresponding bile salt concentration. Using the regression  
103 equation obtained from the established calibration plot of each drug at each bile salt  
104 concentration, the concentration of solubilised drug was determined. A plot of the amount  
105 solubilised with bile salt concentration facilitated calculation of the solubilisation ratio (SR)  
106 whereby the mole fraction solubilised ( $X_m$ ) is equal to  $SR/(1 + SR)$  and can be combined with  
107 the literature-based calculated mole fraction aqueous solubility ( $X_a$ ) to determine the  
108 micelle/water partition coefficient ( $K_{x_m/a}$ ) as follows<sup>14</sup>:

$$109 \quad K_{x_m/a} = X_m / X_a$$

110 Results from the UV analysis permitted the development of a dataset that contained  
111  $\log K_{x_m/a}$  values for 20 compounds along with their physicochemical parameters (e.g.  
112 molecular weight, rotatable bonds, molar volume, number of hydrogen bond acceptors)  
113 published human intestinal absorption (HIA) values facilitating development of an equation  
114 to relate  $\log K_{x_m/a}$  with HIA using simple linear regression in combination with the established  
115 equation:

$$116 \quad \text{Logit HIA} = \log [\% \text{ HIA} / (100 - \% \text{ HIA})] \quad 15$$

117 A further four compounds were then similarly analysed by measuring  $\log K_{x_m/a}$ , to predict  
118 % HIA. A comparison was then made between the predicted values with those published in  
119 literature. Simple linear regression analysis was carried out using Minitab 17<sup>®</sup> (Minitab Inc.,  
120 State college, PA, USA; licensed to the University of Huddersfield) where the previously  
121 mentioned dataset was imported into it. The final model was obtained by excluding molecular  
122 descriptors which were not statistically significant (P-value > 0.05) and those with

123 unacceptably high levels of variance inflation factor (VIF), which is considered as a  
124 multicollinearity indicator, were not included in the final model. Cook's distance and  
125 residuals were used to detect whether any of the model variables had high leverage. The  
126 optimal final model was then obtained including only  $\log K_{xm/a}$  as a predictor for logit HIA,  
127 the model was then validated using four compounds.

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## 129 **Results and Discussion**

130 In total, twenty four drugs were analysed to determine the concentration of drug in  
131 solution as a function of NaDC concentration, these were selected to cover a range of  
132 physicochemical properties, such as reported HIA,  $\log P_{o/w}$  and other properties. All data  
133 were then plotted to determine a SR value for each drug (i.e. the slope): a selection of which  
134 can be seen in Figure 1.

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144 Figure 1 clearly shows a linear relationship between the concentration of drug and the  
145 concentration of NaDC. Only linear sections of the plots were incorporated to calculate SR,  
146 some were deemed to be non-linear, such as the lower concentrations of quinine and the

147 higher concentrations of acetaminophen (data not shown). These non-linear relationships may  
148 be due to preferential drug-drug interactions rather than drug-NaDC interactions as such  
149 drugs are known to self-associate <sup>16</sup>. The majority of the compounds did exhibit a linear  
150 relationship over the concentration range studied (7 – 20 mM) ensuring confidence in the  
151 experimental system. Using the calculated SR value, along with the mole fraction of aqueous  
152 solubility for the drug, facilitated calculation of a micelle/aqueous partition coefficient for  
153 each drug. By analysing these values alongside reported % HIA literature data (Table 1)  
154 enabled the application of simple linear regression to construct an equation to permit  
155 calculation of % HIA for any compound through measurement of its solubility in NaDC.

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165 Using data presented in Table 1, simple linear regression analysis was utilised to create an  
166 optimised equation to predict HIA:

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$$\text{Logit HIA} = -0.919 + 0.4618 \log K_{xm/a}$$



168  $S = 0.236264$   $R^2 = 0.8492$ ,  $R^2_{adj} = 0.8409$ ,  $R^2_{Pred} = 0.8232$ ,  $F = 101.388$ . The P-values obtained  
169 for this model indicate that the relationship between % HIA and  $\log K_{xm/a}$  values was  
170 statistically significant at the 95 % confidence level where they were  $< 0.05$ , also the model's  
171 F-ratio was found to be statistically significant.  $\log K_{xm/a}$  was found to have a 95 %  
172 confidence interval of (0.365, 0.558) and a t-value of (10.069). According to Cook's distance  
173 and residuals, no drug among the model's dataset was found to be influential or having high  
174 leverage. The unadjusted  $R^2$  of 0.8492 derived from the current data indicates that the fit of  
175 the sampled drugs to the model is good. The  $R^2_{Pred}$  value of 0.8232 indicates that the fit of  
176 the drugs to the model is valid and confirms the potential suitability of UV measurement of  
177 solubility using NaDC to predict oral drug absorption in the human GI tract. The close values  
178 of  $R^2_{adj}$  and  $R^2_{Pred}$  show no evidence of the current model over-fitting the data. A Durbin-  
179 Watson statistic value of 2.309 proves the absence of autocorrelation in the current regression  
180 model. A summary of predicted % HIA values using the established equation alongside  
181 published literature values for % HIA can be seen in Table 2 including validation  
182 compounds.

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193 A plot of calculated % HIA with corresponding literature values can be seen in Figure  
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## 203 **Conclusions**

204 Overall, the development of an equation to predict % HIA using a simple UV based  
205 technique via calculation of the micelle/water partition coefficient, has been shown to be  
206 statistically appropriate and reliable as a method to determine intestinal absorption. Using a  
207 simple, economic and robust UV bile salt assay allows prediction of human intestinal  
208 absorption and avoids many of the disadvantages of other techniques, such as animal based  
209 methods.

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## 211 **References**

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213 1. (a) Egan, W. J.; Lauri, G., Prediction of intestinal permeability. *Advanced Drug Delivery*  
214 *Reviews* 2002, 54 (3), 273-289; (b) Votano, J. R.; Parham, M.; Hall, L. H.; Kier, L. B., New predictors  
215 for several ADME/Tox properties: Aqueous solubility, human oral absorption, and Ames  
216 genotoxicity using topological descriptors. *Molecular Diversity* 2004, 8 (4), 379-391.

- 217 2. (a) Basant, N.; Gupta, S.; Singh, K. P., Predicting human intestinal absorption of diverse  
218 chemicals using ensemble learning based QSAR modeling approaches. *Computational Biology and*  
219 *Chemistry* 2016, *61*, 178-196; (b) De Julián-Ortiz, J. V.; Zanni, R.; Gálvez-Llompart, M.; García-  
220 Domenech, R., The prediction of human intestinal absorption based on the molecular structure.  
221 *Current Drug Metabolism* 2014, *15* (4), 380-388.
- 222 3. Kostewicz, E. S.; Aarons, L.; Bergstrand, M.; Bolger, M. B.; Galetin, A.; Hatley, O.; Jamei,  
223 M.; Lloyd, R.; Pepin, X.; Rostami-Hodjegan, A.; Sjögren, E.; Tannergren, C.; Turner, D. B.; Wagner,  
224 C.; Weitschies, W.; Dressman, J., PBPK models for the prediction of in vivo performance of oral  
225 dosage forms. *European Journal of Pharmaceutical Sciences* 2014, *57* (1), 300-321.
- 226 4. Kostewicz, E. S.; Abrahamsson, B.; Brewster, M.; Brouwers, J.; Butler, J.; Carlert, S.;  
227 Dickinson, P. A.; Dressman, J.; Holm, R.; Klein, S.; Mann, J.; McAllister, M.; Minekus, M.; Muenster,  
228 U.; Müllertz, A.; Verwei, M.; Vertzoni, M.; Weitschies, W.; Augustijns, P., In vitro models for the  
229 prediction of in vivo performance of oral dosage forms. *European Journal of Pharmaceutical*  
230 *Sciences* 2014, *57* (1), 342-366.
- 231 5. Waters, L. J.; Shokry, D. S.; Parkes, G. M., Predicting human intestinal absorption in the  
232 presence of bile salt with micellar liquid chromatography. *Biomedical Chromatography* 2016.
- 233 6. Guerra, A.; Denis, S.; le Goff, O.; Sicardi, V.; François, O.; Yao, A. F.; Garrait, G.; Manzi, A.  
234 P.; Beyssac, E.; Alric, M.; Blanquet-Diot, S., Development and validation of a new dynamic  
235 computer-controlled model of the human stomach and small intestine. *Biotechnology and*  
236 *Bioengineering* 2016, *113* (6), 1325-1335.
- 237 7. Augustijns, P.; Wuyts, B.; Hens, B.; Annaert, P.; Butler, J.; Brouwers, J., A review of drug  
238 solubility in human intestinal fluids: Implications for the prediction of oral absorption. *European*  
239 *Journal of Pharmaceutical Sciences* 2014, *57* (1), 322-332.
- 240 8. Moghimipour, E.; Ameri, A.; Handali, S., Absorption-Enhancing Effects of Bile Salts.  
241 *Molecules* 2015, *20* (8), 14451-14473.
- 242 9. (a) Esposito, G.; Giglio, E.; Pavel, N. V.; Zanobi, A., Size and shape of sodium deoxycholate  
243 micellar aggregates. *Journal of Physical Chemistry* 1987, *91* (2), 356-362; (b) Matsuoka, K.; Moroi,  
244 Y., Micelle formation of sodium deoxycholate and sodium ursodeoxycholate (Part 1). *Biochimica et*  
245 *Biophysica Acta - Molecular and Cell Biology of Lipids* 2002, *1580* (2-3), 189-199.
- 246 10. D'Alagni, M.; D'Archivio, A. A.; Galantini, L.; Giglio, E., Structural study of the micellar  
247 aggregates of sodium chenodeoxycholate and sodium deoxycholate. *Langmuir* 1997, *13* (22), 5811-  
248 5815.
- 249 11. Bogdanova, L. R.; Gnezdilov, O. I.; Idiyatullin, B. Z.; Kurbanov, R. K.; Zuev, Y. F.; Us'yarov,  
250 O. G., Micellization in sodium deoxycholate solutions. *Colloid Journal* 2012, *74* (1), 1-6.
- 251 12. Waters, L. J.; Hussain, T.; Parkes, G. M. B., Thermodynamics of micellisation: Sodium  
252 dodecyl sulfate/sodium deoxycholate with polyethylene glycol and model drugs. *The Journal of*  
253 *Chemical Thermodynamics* 2014, *77* (0), 77-81.
- 254 13. Wiedmann, T. S.; Liang, W.; Kamel, L., Solubilization of drugs by physiological mixtures of  
255 bile salts. *Pharmaceutical Research* 2002, *19* (8), 1203-1208.
- 256 14. Wiedmann, T. S.; Kamel, L., Examination of the solubilization of drugs by bile salt micelles.  
257 *Journal of Pharmaceutical Sciences* 2002, *91* (8), 1743-1764.
- 258 15. Akamatsu, M.; Fujikawa, M.; Nakao, K.; Shimizu, R., In silico prediction of human oral  
259 absorption based on QSAR analyses of PAMPA permeability. *Chemistry and Biodiversity* 2009, *6*  
260 (11), 1845-1866.
- 261 16. Casabianca, L. B.; De Dios, A. C., <sup>13</sup>C NMR study of the self-association of chloroquine,  
262 amodiaquine, and quinine. *Journal of Physical Chemistry A* 2004, *108* (40), 8505-8513.

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