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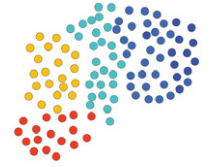
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Interrogation of modern and ancient genomes reveals the complex domestic history of cattle



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Implications

- The analysis of mitochondrial and nuclear DNA sequence polymorphisms from modern cattle populations has had a profound impact on our understanding of the events surrounding the domestication of cattle. From these studies, it has been possible to distinguish between pre- and post-domestic genetic differentiation, supporting previous assertions from archaeological studies and, in some cases, revealing novel aspects of the demographic history of cattle.
- Analyses of genetic material retrieved from the remains of extinct ancestral wild cattle have also added valuable layers of information pertaining to cattle domestic origins; however, information from these investigations have, in general, been limited to small, variable portions of the mitochondrial genome owing to technical challenges associated with the retrieval and amplification of ancient DNA.
- In recent years, however, new high-throughput, massively parallel genomics technology platforms, such as single-nucleotide polymorphism (SNP) genotyping arrays and next-generation sequencing (NGS), have provided a new impetus to the studies of genetic variation in extant and ancient cattle.
- Arrays of SNP have facilitated high-resolution genetic surveys of global cattle populations and detection of ancient and recent genomic selective sweeps. Next-generation sequencing analyses of modern and ancient cattle hold great promise for identifying and cataloging of pre- and post-domestication patterns of genomic variation and correlating this with natural and artificial selection processes.

Key words: cattle domestication, mitochondrial DNA, next-generation sequencing, selection, single-nucleotide polymorphism

Introduction

The development of agriculture at the advent of the Neolithic period, *circa* 13,000 years before present (YBP), is considered to be the most important cultural innovation in human history. Before this, anatomically modern humans had successfully managed to occupy most habitable and accessible regions of the world as small groups of widely dispersed hunter-gatherers, exploiting a diverse range of wild plant and animal species. The transition in human subsistence from nomadic forager to sedentary agriculturist had a number of profound cultural, economic, and demographic consequences. Through the cultivation of crops and the domestication of large animals, humans could manipulate and manage their natural environment in a completely new way. The concomitant increase in food and other biological resources facilitated a significant increase in human population numbers, leading to the geographic expansion of agriculturalist communities and the eventual emergence of complex urban societies. Consequently, deciphering the anthropological and biological processes associated with plant and animal domestication is key to understanding the origins of modern human society (Diamond, 2002).

Domestication has been defined as “*the human creation of a new form of plant or animal—one that is identifiably different from its wild ancestor and extant wild relatives*” (Smith, 1995); in terms of animal domestication, this can be viewed as an elaboration of the predator–prey relationships between *Homo sapiens* and target species. Animal domestication is generally a long-term cumulative process, characterized by gradual changes in the behavior of both human and animal populations over time, which has had selective advantages for both biological partners and has been aptly described as a “covenant of the wild” (Budiansky, 1994). As with plants, there are two key elements associated with the initial domestication of animals: at first, it is most likely that humans segregated a few individuals from wild populations; following this, active intervention in the life cycles of captive populations lead to a behavioral and physiological trajectory toward full domestication. This was not an instantaneous event in which a wild animal was immediately transformed into a domesticate; instead it was a process that required sustained breeding of captured and tamed animals over prolonged periods of time, with the change from wild to domestic forms occurring after several domestic generations since initial capture from the wild (Uerpmann, 1996). Sustained human management of early domestic populations led to an increase in animal numbers and an expansion of species’ geographic range beyond that of



their wild progenitors. From the perspective of these early human herders, this led to security and conservation of animal food supplies, especially during the periods of environmental and climatic unpredictability that coincided with the advent of agriculture (Diamond, 2002; Bar-Yosef, 2011).

Zooarchaeological Investigation of Animal Domestication

Much of what is known about the ‘when’ and ‘where’ of animal domestication has been gleaned from the scientific examination of faunal material retrieved from archaeological sites. These zooarchaeological studies of animal domestication are largely based on osteological analysis and radiocarbon dating of faunal remains at early farming settlements. Geographic centers of animal domestication are those where, over time, a staged, temporal transition from characteristically wild to domestic faunal patterns are evident in response to changes in human behavior (Meadow,

1989). In this regard, many of the biological changes that occur due to domestication are morphological, and these observable changes allow archaeologists to ascertain and catalog evidence for animal domestication at a particular site (Clutton-Brock, 2012).

It is thought that selection for docility and reduced aggression in wild and early domesticates were indirectly associated with gradual, yet distinctive, changes in secondary morphological characters, such as the form and fine structure of bones. These include the shortening of the skull that resulted in the crowding and reduced size of teeth, reductions in brain size, and changes in horn morphology (Bökönyi, 1976; Smith, 1995). Changes in morphological markers are generally harder to distinguish in early domesticates; for example, Bökönyi (1976) considers that well-defined morphological changes do not occur for about 30 generations (approximately 200 years in the case of cattle). Nevertheless, at least two kinds of morphological change are thought to have occurred relatively early in the domestication process. The first is the appearance of bone pathologies, reflecting the physical trauma (caused, for example, by the use of the domesticate for draught), poor diet, and higher infection rates resulting from enforced confinement; the second is a decrease in size due to selection of more manageable animals and malnutrition (Bökönyi, 1976; Smith, 1995).

Additional evidence includes temporal changes in the proportions of age and sex groups, whereby the demography of faunal assemblages produced by human hunting differs from those produced from managed herds. This may suggest a pre-selection of animals for domestication (for example, young female animals may have been more easily captured and tamed than wild males) or differential use of sex groups (for example, young males may have been preferentially slaughtered for food rather than reproductively active females). Finally, ancillary archaeological evidence for animal domestication includes artistic representations (rock art

Box 1. Genetics and Domestication

The principles of Darwinian evolution predict that modern populations share biological (and therefore genetic) continuity with past populations of similar geographical provenance. Therefore, modern patterns of genetic diversity reflect signatures of past demographic events. Previous population expansions, contractions (or “bottlenecks”), migrations (to and from populations with subsequent breeding), amalgamations, and selection events (natural or artificial) leave characteristic signatures on genomes in the form of altered allele frequencies, haplotype structures, and selective sweeps, which are transmitted to succeeding generations. The extent of genetic differences between populations or species is indicative of their relationship since they shared a common ancestor. Therefore, modern genomes can be viewed as “molecular heirlooms” that have been transmitted from past to present generations and from which past population histories can be reconstructed. When an ancestral population splits, the frequencies of alleles represented within each separate group will slowly change over time due to the processes of mutation, random genetic drift, and selection. It is these differences that can be assessed to determine the time depth of the divergence event.

Consequently, studies of molecular genetic variation provide a more sensitive tool with which to assess breed and population relationships than measurements of morphological characters.

Investigations of domestic origins and genetic diversity in modern populations can be viewed as an extension of the approach conceived by Nikolai Vavilov in the 1920s (Vavilov, 1926). He realized that elevated phenotypic (and genetic) diversity within a defined geographic area, together with an overlapping distribution between domestic and pre-domestic forms, represents biological evidence for domestication. Accordingly, modern animals originating from a purported domestication center are expected to display elevated levels of genetic diversity due to retention of more wild ancestral variation compared with recently derived populations where a proportion of ancestral variation has been lost through subsampling during successive phases of migration from these centers. Furthermore, as there is a correlation between time and mutation, older populations are expected to have accumulated more DNA sequence variation over time compared with younger populations; therefore, direct modern descendants of the early domesticates should exhibit increased genetic variation as they have had more time to accumulate new mutations compared with younger derived populations.

and figurines) depicting captured wild and domestic animals (Zeder et al., 2006).

The Domestic Taxa of Cattle

Cattle are the most economically important of domesticated animal species and number more than 1300 million on a global scale, constituting some 800 extant cattle breeds (FAO, 2012). Domestic cattle are classified with binomial Linnean names, following convention and the work of Baker and Manwell (1980) and Grigson (1980): *Bos taurus* denotes humpless taurine cattle and *Bos indicus* denotes humped zebu (or *indicine*) cattle.

Taurine cattle predominate in northern and western Africa and almost the whole of the Eurasian landmass, from northwest Europe to Japan, whereas zebu cattle are native to the Indian subcontinent and are also found in the Near and Middle East and most of eastern and southern Africa.

Zebu cattle differ from taurine cattle in a number of significant aspects: the presence of a muscular fatty hump of variable size positioned either on the shoulders (cervico-thoracic) or the neck (cervical); a larger, more pendulous dewlap; drooping ears; a naval flap; and tolerance of arid environments. Zebu cattle also display a lower basal metabolic rate, lower water and nutrient requirements, and are generally more resistant to ticks and intestinal parasites than taurine animals. However, unlike zebu cattle, indigenous African taurine cattle display an inherited tolerance to the symptoms of trypanosomiasis, a wasting disease caused by flagellate protozoans of the genus *Trypanosoma* and transmitted by tsetse flies (*Glossina* sp.; Payne, 1991).

Despite differences in morphology and disease tolerance/susceptibility, both domestic taxa possess the same number of chromosomes ($2n = 60$; Melander, 1959). With regard to the sex chromosomes, there is one form of the X chromosome in cattle, which is submetacentric in both taurine and zebu cattle. In contrast, there are two forms of the Y chromosome, with the typical *B. taurus* Y chromosome morphology being submetacentric and *B. indicus* having small acrocentric Y chromosomes. As taurine and zebu cattle can freely interbreed, producing fertile offspring, they are often regarded as subspecies within the *Bos* genus (Bökönyi, 1997).

The Domestic Origins of Cattle

It is now widely accepted that the two modern domestic cattle taxa were derived from the now-extinct wild ox, or aurochs (*Bos primigenius*). *Bos primigenius* evolved during the interglacial period of the mid-Pleistocene about 500,000 YBP (Guintard, 1999) and are thought to be descendants of *B. acutifrons* (Pilgrim, 1947). During the Middle and Upper Pleistocene, aurochs were widespread throughout Eurasia, and through differences in horn shape and body size, three continental races have been identified: *B. primigenius primigenius* (Eurasia), *B. p. namadicus* (South



Asia), and *B. p. opisthonomus* [North Africa] (Grigson, 1978, 1980). In Eurasia, the range of the aurochs expanded and contracted in response to interglacial and glacial stages during the Ice Age. After the last deglaciation, around 12,000 YBP, aurochs populated much of Eurasia and portions of northern Africa, stretching from Britain to China, with the exception of northern Scandinavia, Ireland, and Siberia.

With a derivation from the Greek word *oros*, meaning mountain, the word aurochs, translated from German, is both singular and plural and literally means “primeval ox” or “proto-ox.” Aurochs bulls were pronouncedly larger than modern domesticates, up to 2 m at the shoulder, and were often equipped with extremely long horns (Zeuner, 1963). Although the last surviving European aurochs reportedly died in a Polish animal reserve in AD 1627 (Zeuner, 1963), zooarchaeological analysis and contemporary descriptions of this animal suggest that the domestication of wild cattle was a considerable achievement. Indeed, Julius Caesar, in his record of the Gallic Wars, wrote of them: “They are a little below the elephant in size and of the appearance, color, and shape of a bull. Their strength and speed are extraordinary; they spare neither man nor wild beast which they have espied....But not even when taken very young can they be rendered familiar to men and tamed.”

The earliest recorded archaeological evidence for cattle domestication comes from the early Neolithic site of Dja'de el Mughara in the Middle Euphrates Valley in southwest Asia (Helmer et al., 2005). Progressive reductions in the sizes of fossil cattle bones through successive depositional layers have been documented, dating to 10,750–10,250 calibrated (cal.) YBP, and these are thought to be representative of the first domestic *B. taurus* cattle derived from the Eurasian aurochs, *B. p. primigenius*. There is also morphological evidence for management of taurine cattle in northeastern China at this time (10,660 cal. YBP), which may indicate local domestication of autochthonous aurochs (Zhang et al., 2013).

It is generally accepted that taurine cattle, along with other Near Eastern domesticates, such as sheep, goats, barley, and wheat, spread into Europe via two routes: a *Mediterranean* route and a *Danubian* (or *Continental*) route. Archaeological evidence suggests the dispersal of agriculture in Europe began around 9,000 cal. YBP, with the first European farming settlement located at Argissa Magoula in Greece at 8,600 cal. YBP (Rein-

Box 2. mtDNA as a Marker for Population Genetics

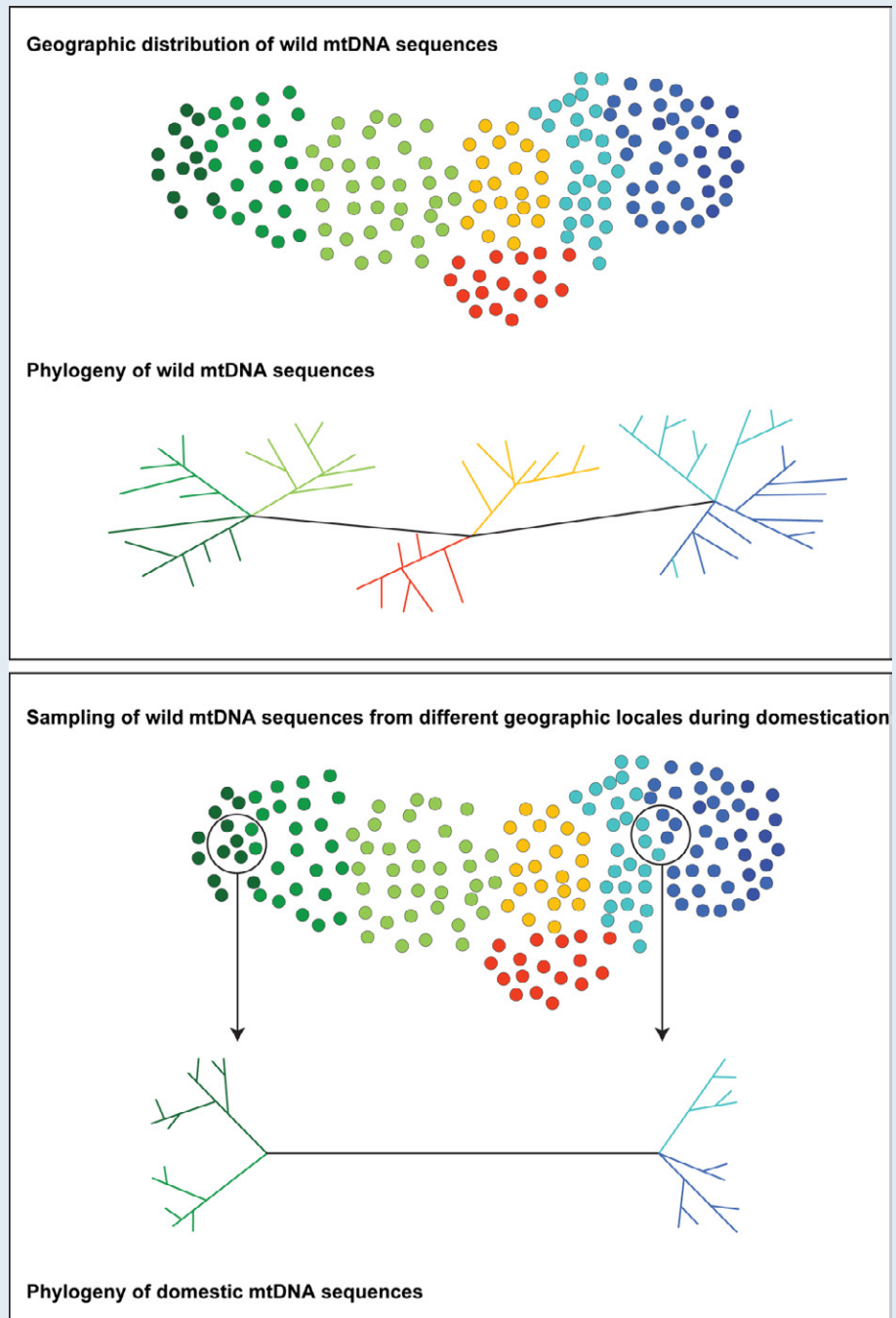
Different classes of DNA markers have been used to reconstruct the ancestry and domestic history of cattle. Mitochondrial DNA (mtDNA) has been extensively used to test hypotheses about the domestication of animals. Mammalian mitochondrial chromosomes are small, circular organellar genomes (~16,300 bp) that display a maternal mode of inheritance, an absence of recombination, and a rapid rate of mutation relative to the nuclear genome. Thus, mtDNA lineages are regarded as clonal, information-rich, singly inherited units (or haplotypes) from which matriarchal evolutionary histories can be reconstructed for domestic populations that otherwise may have complex genetic histories.

Typically, phylogenies of wild ungulate mtDNA show multiple clades and a complex branching structure resulting from a wide geographic sampling of wild variability (Bradley and Maguee, 2006). In contrast, mtDNA phylogenies constructed from modern cattle show two major distinct clades, representative of *B. taurus* and *B. indicus* sequences. As domestication results in the attenuated, geographically restricted sampling of wild variation, the two major mtDNA clades found in domestic cattle correspond to the mtDNA sequences sampled from the Eurasian and South Asian aurochs landraces during the domestication process. The extant diversity within the *B. taurus* and *B. indicus* clades also contain the limited amount of diversity that has accrued via mutation within the ~10,000-year time frame of cattle domestication (Figure 1B).

The consequence of domestication for mtDNA sequence diversity is illustrated in the figure below where phylogenetically distinct mtDNA sequences (colored circles, in which similar mtDNA sequences show similar shading) sampled from wild extant populations across wide geographic locales generate “bushy” phylogenies consisting of multiple clades with complex branching structures. In contrast, the attenuated sampling of mtDNA sequence variation from geographically and genetically disparate wild populations causes sequences to cluster into small numbers of clades, often generating “double-headed broomstick” topologies, such as that observed for *B. taurus* and *B. indicus* (MacHugh and Bradley, 2001)

Studies have also shown that for domestic mammalian livestock, mtDNA sequences show geographic inertia due to maternal inheritance and low migration of females under managed breeding (i.e., modern domestic animals sampled from similar locations retain distinct mtDNA haplotypes despite millennia of interbreeding with animals, usually males, imported from

other locations). Consequently, sequences that originate from different captures of a diverse wild species can maintain a phylogenetic distinction even after millennia of predominantly male-mediated domestic inbreeding, which allows the resolution of pre-domestic patterns of diversity (for example, the sequence divergence between modern *B. taurus* and *B. indicus*) and post-domestic patterns of sequence diversity (for example, diversity within each of the major taurine and zebu haplogroups) to be discerned. The majority of bovine mtDNA phylogeographic studies have targeted the hypervariable mtDNA control region or displacement loop (D-loop); however, more recent studies have analyzed the complete mitochondrial genome.



Box 3. Y-chromosomal and Autosomal Genetic Markers for Population Genetics

Evolutionary history of male lineages can be examined using polymorphic markers distributed across the paternally inherited Y chromosome. Y chromosomal markers largely consist of simple tandem repeats (STR), such as microsatellites, which are tandemly repeated short non-coding DNA sequences (for example, CACA-CACACACACACA). These repetitive arrays are often highly polymorphic in length due to variation in the number of repeat units, which generates a range of alleles that are transmitted in a Mendelian fashion (Underhill and Kivisild, 2007). The STR are ubiquitously distributed throughout the mammalian nuclear genome and can also be used to analyze genetic variation in the autosomal genome exclusive of the Y chromosome. Generally, autosomal and Y chromosomal STR marker length variation is presumed to be selectively neutral, unless the locus under examination is closely linked to a selected coding region (Rosenbaum and Deinard, 1998).

More recently, whole-genome DNA sequencing has revealed the abundance of SNP that exist in eukaryotic nuclear genomes. These are defined as single base pair substitutions in genomic DNA, where the least common allele has a frequency of ≥ 0.01 in a particular population (Kruglyak and Nickerson, 2001). On average, SNP occur once every 500 to 1,000 bp of mammalian DNA sequenced and are found in both coding and non-coding nuclear DNA (Bovine Genome Sequencing and Analysis Consortium et al., 2009). Although the vast majority of SNP in mammalian genomes are biallelic (owing to their low mutation rate) and therefore have reduced information content compared with STR, their pan-genomic distribution and digital nature (where a SNP locus is queried for the presence or absence of an allele) has facilitated development of high-throughput genotyping arrays or “SNP chips” (Matukumalli et al., 2009). This has resulted in their widespread use in more recent population genetic investigations in cattle and other domestic livestock.

gruber and Thissen, 2009). Farming then moved into the plains of south-eastern Europe between 7,850 and 7,450 cal. YBP (Dolukhanov et al., 2005), reaching Portugal around 7,250 cal. YBP (Waterbolk, 1968; Zilhao, 2001; Tresset and Vigne, 2007). Migration along a continental route into Poland and Germany occurred around 7,650 cal. YBP, with an expansion of the Bandkeramik culture into northern Europe at 7,250–6,850 cal. YBP (Dolukhanov et al., 2005). The Neolithic period reached northwestern France at 7,150 cal. YBP, and southern Scandinavia, the British Isles, and Ireland were reached after 6,000 cal. YBP (Waterbolk, 1968; Zilhao, 2001; Tresset and Vigne, 2007).

In South Asia, bones of *B. indicus* have been found at Neolithic sites of the Indus Valley (present-day Pakistan and northwest India) dating to circa 7,000 YBP (Meadow, 1993). These sites represent a candidate location for an eastern center of domestication of *B. indicus* cattle from the South Asian aurochs, *B. primigenius namadicus* (Grigson, 1980). Some authors have also argued for the domestication of *B. indicus* within southern India circa 5,000 YBP, citing faunal evidence retrieved from the sites of Piklihal, Utnur, and Maski (Allchin and Allchin, 1968; Allchin and Allchin, 1974; Misra, 2001).

Although archaeological evidence suggests that the earliest domestic cattle of the African continent were humpless *B. taurus*, and were present in North Africa, the eastern Sahara, and near the Nubian Nile by 7,550–6,950 cal. YBP (Wendorf and Schild, 2001), their origins remain controversial. Traditionally, it was held that *B. taurus* was introduced into Africa via the Isthmus of Suez from the Near East approximately 9,000–7,000 YBP (Epstein and Mason, 1984); however, some authors contend that the founding *B. taurus* of Africa was derived from an independent domestication of the indigenous aurochs, *B. p. opisthonotus*, in the eastern Sahara 9,500–8,900 cal. YBP (Wendorf and Schild, 1994; Grigson, 2000; Wendorf and Schild, 2001; Marshall and Hildebrand, 2002).

Bos indicus cattle were initially thought to have entered sub-Saharan Africa via the Horn of Africa from southwest Asia between 700 and 1500 AD, before and during Arab migrations (Epstein and Mason, 1984; Hourani, 1991). However, evidence from Ngamuriak, a Neolithic site in Kenya, suggests that these cattle breeds were present in East Africa at least 1,500 years earlier (Marshall, 1989). Thereafter, exogenous zebu and native taurine cattle were crossbred to form populations of hybrid *sanga* cattle, the modern descendants of which are found in southern and eastern Africa. Internal African pastoralist migrations are believed to have been responsible for dispersals of *sanga* cattle into southern and central African regions (Epstein and Mason, 1984).

Genetic Signatures of Animal Domestication

The terms “wild” and “domestic” suggest a clear dichotomy between forms and do not consider the slow gradual transition in forms that occurred as human–animal relationships intensified during the domestication process. In contrast, geneticists investigating animal domestication do so within the context of a clearly defined process: the genetic response to the managed breeding of animals. As genetic information is transmitted to succeeding generations, modern populations carry in their genome the signatures of past demographic processes and the evolutionary forces that have shaped these. Consequently, all modern domesticates have descended, with modification, from the wild animals that were incorporated into a finite genetic pool at various stages within the time frame of domestication. Modern domestic populations, therefore, are at the endpoint of a temporal genetic continuum that stretches from the initial phases of animal domestication and includes only those influential interactions between human and animals that have left a genetic legacy (Box 1).

Geneticists examining the origins and ancestry of domestic animals focus largely on the analysis of evolutionarily neutral, non-coding loci of the nuclear genome, for example, autosomal and Y chromosome simple tandem repeat (STR) and single-nucleotide polymorphisms (SNP), and extranuclear genomes, such as the mitochondrial genome (mtDNA). These alternate genetic markers, which display different modes of inheritance, information content, and rates of mutation, have been used to unravel the domestic ancestry of cattle (Boxes 2 and 3). More recently, high-throughput genomic technologies such as SNP genotyping array platforms and whole-genome sequences from animals of wide geographic provenance have enabled high-resolution analysis of the genetic relationships among breeds and have identified discrete segments of cattle genomes that have been subject to the selection processes that underlie domestication.

Mitochondrial DNA Genetic Data Support the Independent Domestic Origins of *B. taurus* and *B. indicus*

Wide-ranging surveys of genetic variation in modern cattle populations have been instrumental in showing that all domesticated cattle derive from either taurine or zebu wild progenitors or are hybrids of the two domestic taxa. The most striking aspect of these studies has been the repeated support for the deep divergence between modern *B. taurus* samples from Africa and Europe and *B. indicus* animals from India (Fig. 1A-C). This is illustrated by the high number of nucleotide differences between taurine and zebu mtDNA sequences (Fig. 1A), which in turn, give rise to the long internal branch that separates the *B. taurus* and *B. indicus* clades in the phylogeny shown in Fig. 1B. The extent of sequence divergence between the taurine and zebu mtDNA sequence groups has been estimated in the order of 200,000 years. These patterns of mtDNA diversity indicate the derivation of *B. taurus* and *B. indicus* from at least two geographically separate and genetically divergent wild aurochs populations, presumably the Near Eastern and Asiatic aurochs landraces, respectively, as suggested from zooarchaeological evidence (Loftus et al., 1994a,b).

Notably, genetic diversity within the major taurine and zebu phylogenetic mtDNA sequence groups is markedly (~10 times) smaller than that between them (Achilli et al., 2009). This is illustrated by the small number of nucleotide differences within the taurine and zebu mtDNA sequence groups (Fig. 1A) that generate the short-length branches within each clade (Fig. 1B). As natural populations are expected to display geographic and genetic correlation (such that individuals of similar geographic location displayed increased related genetic variation), the patterns of diversity within each bovine mtDNA sequence clade are consistent with the attenuated sampling (that was limited in time and space) of a subset of aurochs variation concomitant with the domestication process. The extant mtDNA sequence diversity within each clade also includes the limited variation that has accrued through mutation post-domestication (Box 2).

The clear genetic distinction between the taurine and zebu mitochondrial genomes can be used to detect regions where historical gene flow and introgression between the two taxa have occurred since the Neolithic. For example, several studies of Near and Middle Eastern populations have reported zebu mtDNA sequences from samples that are phenotypically and morphologically taurine (Troy et al., 2001; Edwards et al., 2007b; Kantanen et al., 2009). While these populations have appreciable frequencies of zebu mtDNA haplotypes, the quantitative divergence between the taurine and zebu haplotypes remain. This has allowed these phylogeographical exceptions to be securely identified as secondary introductions, which is most likely a legacy of the westward introduction of zebu cattle and subsequent mating with native taurine populations (Troy et al., 2001; Edwards et al., 2007a). With some symmetry, the modern zebu breeds of northern India have been shown to possess a minority component of taurine mtDNA ancestry (Magee et al., 2007; Lenstra et al., 2014). Reciprocal migrations of zebu and taurine cattle from their respective geographic centers of origin, and subsequent mating with native populations elsewhere, were presumably a consequence of the extensive ancient trading routes across southwest Asia, which would have provided an opportunity for genetic exchange between the two taxa (Zeuner, 1963; Naik, 1978; Misra, 2001). In support, available archaeological evidence points towards the presence of zebu cattle in Jordan circa 3,400 YBP and taurine

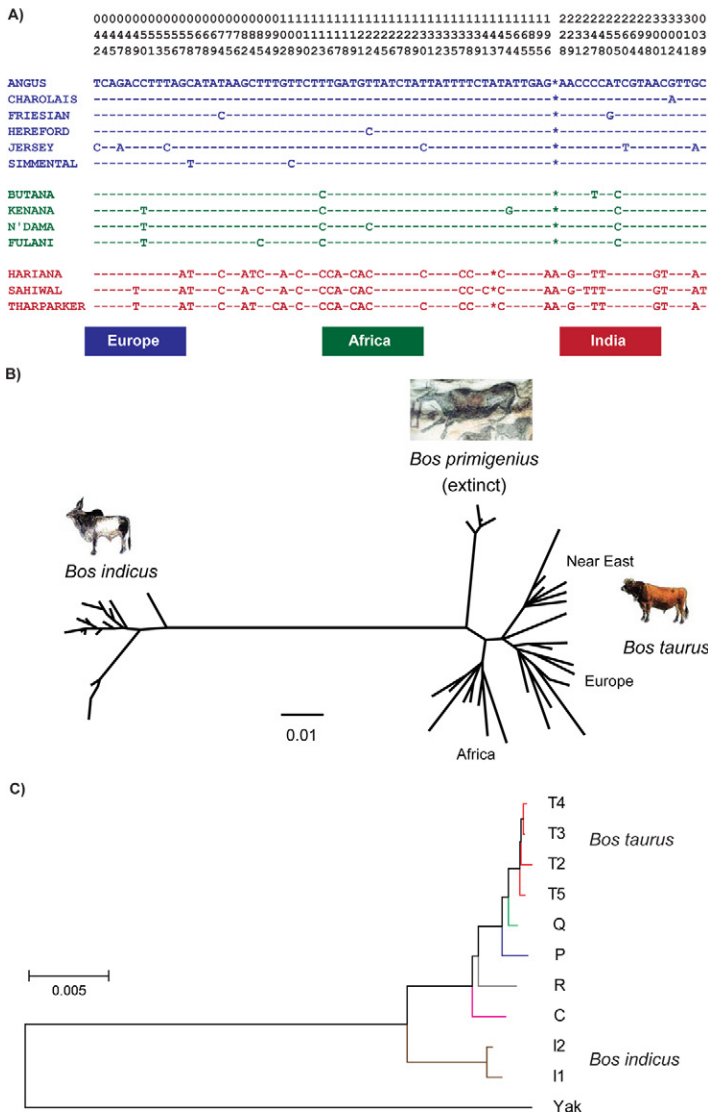


Figure 1A. Alignment of partial mtDNA control region sequences sampled from modern European taurine; African taurine (N'Dama), sanga (Fulani) and zebu (Butana and Kenana); and Indian zebu populations. Only variable sites are shown. Nucleotide substitutions are given; asterisks (*) denote nucleotide insertion/deletions; dashes (-) denote nucleotide identity. The position of the nucleotide differences within the mtDNA control region are reported less 16,000. **Figure 1B.** Neighbor-joining phylogeny constructed from modern *B. taurus*, *B. indicus* and extinct *B. primigenius* mtDNA control-region haplotypes. Analysis is based on a 201 bp fragment for the hypervariable bovine mtDNA control region. The quantitative divergence between modern *B. taurus* and *B. indicus* supports the independent origins of the two domestic taxa. The limited diversity within each sequence group is consistent with the attenuated sampling of diverse wild matrilineal lineages within each domestication center during the domestication process together with the diversity that has accrued since that time (Box 2). The ancient British *B. primigenius* sequences cluster tightly and form an outgroup to the modern *B. taurus* haplotypes, suggesting that there was no major maternal contribution to the European domestic cattle genetic pool from locally recruited wild cattle. Figure adapted from Troy et al. (2001). **Figure 1C.** Neighbor-joining phylogeny of the major bovine mtDNA haplogroups reported in modern and ancient samples based on complete mtDNA genome sequences (16,338 bp). Modern *B. taurus* predominantly possess the T, T1, T2, T3 and T4 haplogroups, while modern *B. indicus* possess haplogroups I1 and I2. Haplogroup P is the major haplogroup from the extinct Eurasian aurochs. Haplogroups Q and R have been recently reported in a small number of modern Italian samples and haplogroup C has been recently reported in an ancient Chinese domestic sample; these data support minor local captures of wild matrilineal lineages. Yak mtDNA is included as an outgroup species.

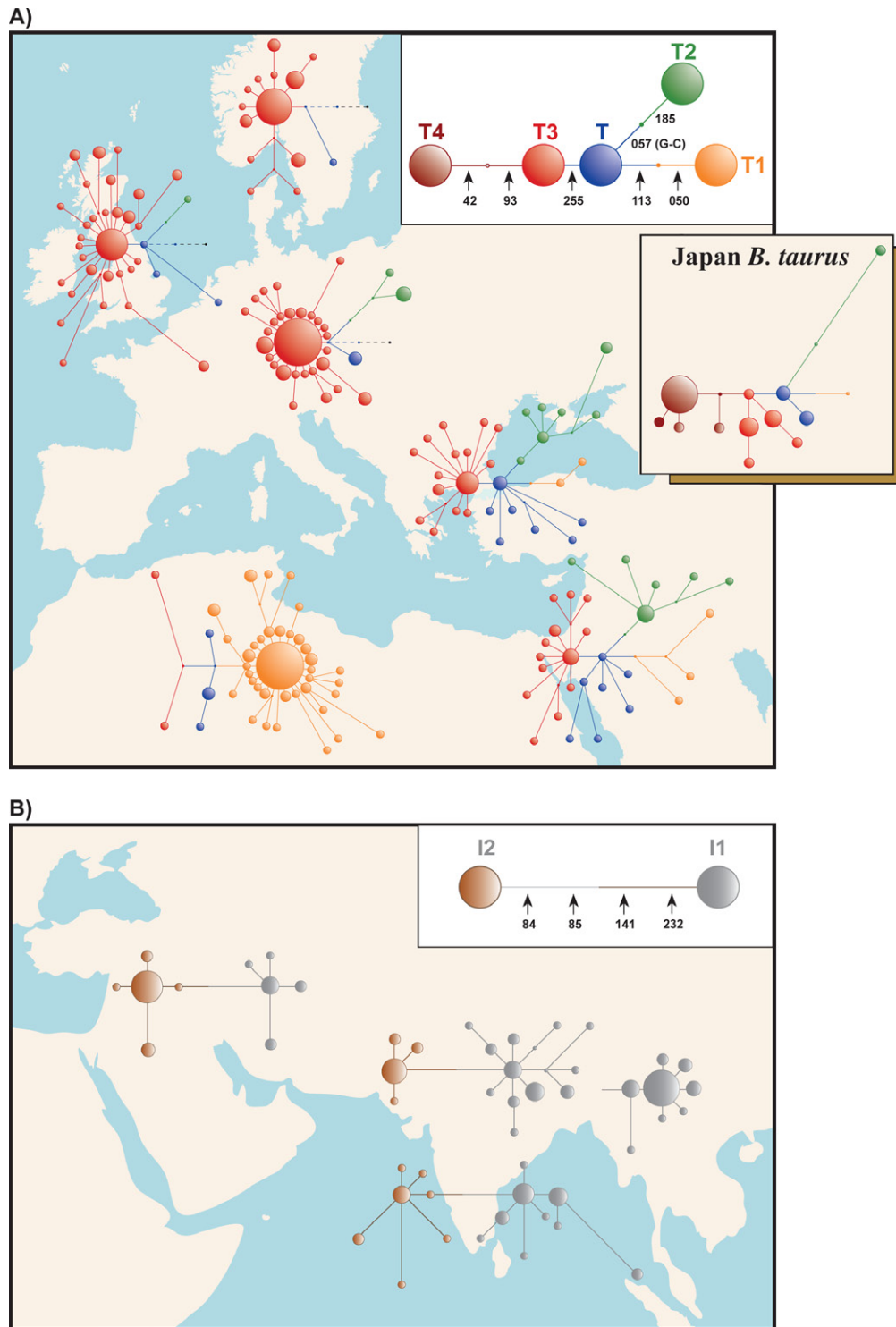


Figure 2. Reduced median network analysis of the most variable 240 bp region of the bovine mtDNA control region shows the regional differences in diversity within modern domestic *B. taurus* (A) and *B. indicus* (B) cattle, superimposed on a map of regions of origin. *B. taurus* and *B. indicus* sequences predominantly fall into five and two major haplogroups, respectively. The positions of the nucleotide substitutions (+16,000) that distinguish each of the major taurine and zebu haplogroups are shown in the skeleton network in the (upper inset in both diagrams). The haplotypes encountered in each region (shaded circles) and the haplogroup to which these haplotypes belong are denoted by the color scheme. Unsamplered intermediate nodes or unsampled primary haplotypes (small points) are also shown. Circle areas are proportional to the number of individuals possessing each mtDNA haplotype. The star-like phylogeny surrounding each of the major taurine and zebu mtDNA centrally-positioned, numerically-predominant and presumed ancestral haplotypes is consistent with past demographic expansions associated with the domestication process. The predominant European haplogroup, T3 (shown in red), is a subset of the diversity encountered in the Near East, where haplogroups T (blue), T2 (green) and T3 are encountered at appreciable frequencies), suggesting that European taurine maternal lineages owe their ancestry to primary domestication centers proximal to the Fertile Crescent rather than to local input from the wild. Both African and Far Eastern populations display two additional clusters, T1 (yellow) and T4 (magenta), which is suggestive of matrilineal input from local wild oxen. In South Asia, zebu sequences fall into either the I1 (grey) or the I2 (brown) haplogroups. Figure adapted from Troy et al. (2001) and Magee et al. (2007)

cattle in the Neolithic settlements of the Indus Valley circa 4,200 YBP (Clason, 1978; Naik, 1978).

Mitochondrial DNA Sequence Diversity Reveals the Complex Ancestry within *B. taurus*

In-depth surveys of partial and complete mtDNA sequence diversity have uncovered more fine-scale aspects of *B. taurus* ancestry. This is demonstrated by the reduced median phylogenetic network analyses (Bandelt et al., 1995) presented in Figure 2. In these, the nucleotide sequence of the most variable 240 bp portion of the bovine mtDNA control region from each sample is aligned to a reference bovine mtDNA genome sequence. The number and nucleotide position of each nucleotide difference from the reference sequence are cataloged for each sample. The data are then displayed visually: distinct mtDNA sequences (or haplotypes) are represented as circles, the areas of which are proportional to the number of samples sharing the same sequence. Nucleotide substitutions, insertions and deletions (i.e., point mutations) that differentiate mtDNA sequences, are represented by lines; the length of each line is proportional to the number of point mutations. Sequences of mtDNA that cluster with each other (based on sequence similarity) form sequence groups or haplogroups.

Inspection of the reduced median networks in Figure 2 reveals that the majority of modern taurine mtDNA sequences fall into one of five phylogenetically distinct, yet closely related, star-like haplogroups, termed T, T1, T2, T3, and T4. Each haplogroup consists of a numerically predominant, centrally positioned haplotype (also termed T, T1, T2, T3 and T4), through which all derived haplotypes root or “coalesce” (Mannen et al., 1998; Troy et al., 2001). A sixth haplotype, T5, has also been recently reported in a small number of modern Italian animals (Achilli et al., 2008, 2009). The numerical and topological predominance of these main haplotypes suggest that they are ancestral sequences incorporated into the domestic pool as a result of the capture and taming of wild female progenitors during the domestication process and which have survived in modern populations. The associated sub-haplotypes are thought to be derived from the ancestral sequence due to the accumulation of mutations since domestication. The star-like patterns of diversity observed within each haplogroup represent the phylogeny expected from a population that has undergone a domestication-induced demographic expansion from a small base (Figure 2). This model proposes that a relatively small number of wild female animals were incorporated into early domestic flocks, which gave rise to increasing numbers of offspring (and consequently, increased mtDNA lineage survivorship) under continuously improving systems of animal management (Bradley and Magee, 2006; Bollongino and Burger, 2007).

Another noteworthy feature of *B. taurus* mtDNA sequence diversity is the spatial distribution of the major haplogroups (Figure 3). In particular, one haplogroup (T3) predominates in Europe and is one of four haplogroups (T, T1, T2, and T3) detected in the Near East at varying frequencies. Haplogroup T2 has appreciable frequencies in Italian, Balkan, and Asian taurine cattle but is found only sporadically in the taurine cattle of central, western, and northern Europe. Also, taurine mtDNA sequence diversity levels are highest in the Near East with reduced diversity observed outside this region. Vavilov’s principle (Vavilov, 1926) is evident in these observations, in which he predicted more ancestral biological variation would be retained at a center of origin for a domestic species. In modern genetic terms, loci in these areas should exhibit higher levels of haplotypic

and nucleotide diversity. Lineages would then be lost through successive colonization events, resulting in reduced diversity in derived populations at a distance from the source. The geographical distribution of taurine mtDNA haplogroups, whereby European diversity is a subsample of the diversity encountered in the Near East, supports the hypothesis that European cattle are a derivative of the Near Eastern Neolithic complex (Troy et al., 2001; Lenstra et al., 2014). This is supported by archaeological evidence and indicates that *B. taurus* domesticates migrated from the Near East with nomadic pastoralists into central and northern Europe following the Danubian route, finally reaching western Europe and Britain around 6,000 cal. YBP (Zilhao, 2001; Brown, 2007).

Haplogroup T1 almost exclusively describes African mtDNA diversity and is found only at low frequencies elsewhere; in Africa, non-T1 haplogroups have been reported only in Egyptian and northwest African populations (Troy et al., 2001; Lenstra et al., 2014). The over-representation of the T1 haplogroup in Africa has been cited as evidence for the separate domestication of the founding populations of the African continent, as previously proposed by archaeological data (Stock and Gifford-Gonzalez, 2013). The near-exclusive predominance of the T1 haplogroup in Africa has enabled geographical exceptions to be identified as secondary migrations from Africa. For example, the detection of African T1 haplogroups in Iberia and Sicily and southern Italy suggests African genetic influence into southern Europe via the Mediterranean Sea coastline (Cymbron et al., 1999; Anderung et al., 2005; Beja-Pereira et al., 2006).

T4 is a subgroup of the T3 haplogroup that has been found in the taurine populations of East Asia (including eastern Siberian, Chinese, Korean, and Japanese cattle) but has not been reported in western Eurasian populations (Figure 2). This has led to some speculation for either another possible domestication center in the Far East or the recruitment of wild matriline into early domestic herds migrating eastwards from the Near East (Mannen et al., 1998; Lai et al., 2006; Kantanen et al., 2009).

Notably, the quantitative difference between the five major taurine ancestral haplotypes is small, with a coalescence time around the central T haplotype estimated at ~16,000 YBP (Troy et al., 2001; Achilli et al., 2008; Ho et al., 2008; Achilli et al., 2009). Thus, the shallow mtDNA sequence divergence observed in modern taurine populations suggests that the population of wild aurochs from which *B. taurus* was derived was itself limited in diversity.

Mitochondrial DNA Sequence Diversity Supports a South Asian Derived Origin of *B. indicus*

A complex picture of *B. indicus* domestication has also emerged from mtDNA studies. Similarly to taurine mtDNA, zebu haplotype sequences fall into two major haplogroups (I1 and I2), comprised of two ancestral haplotypes (also called I1 and I2), through which all other sequences in the phylogeny root (Figures 1C and 2). Comparable to taurine mtDNA variation, the dual star cluster motif within these clusters suggests that they are each signatures of domestication-induced population expansion; the estimated time-depths of the expansions are compatible with zebu domestic history as determined from archaeological studies (Ho et al., 2008). The I1 and I2 haplogroups display some geographic distribution across south Asia, with haplogroup I1 predominating in cattle that have moved eastwards to Southeast Asia from the Indus Valley, the most likely center of origin for this haplogroup. Haplogroup I2 is encountered almost

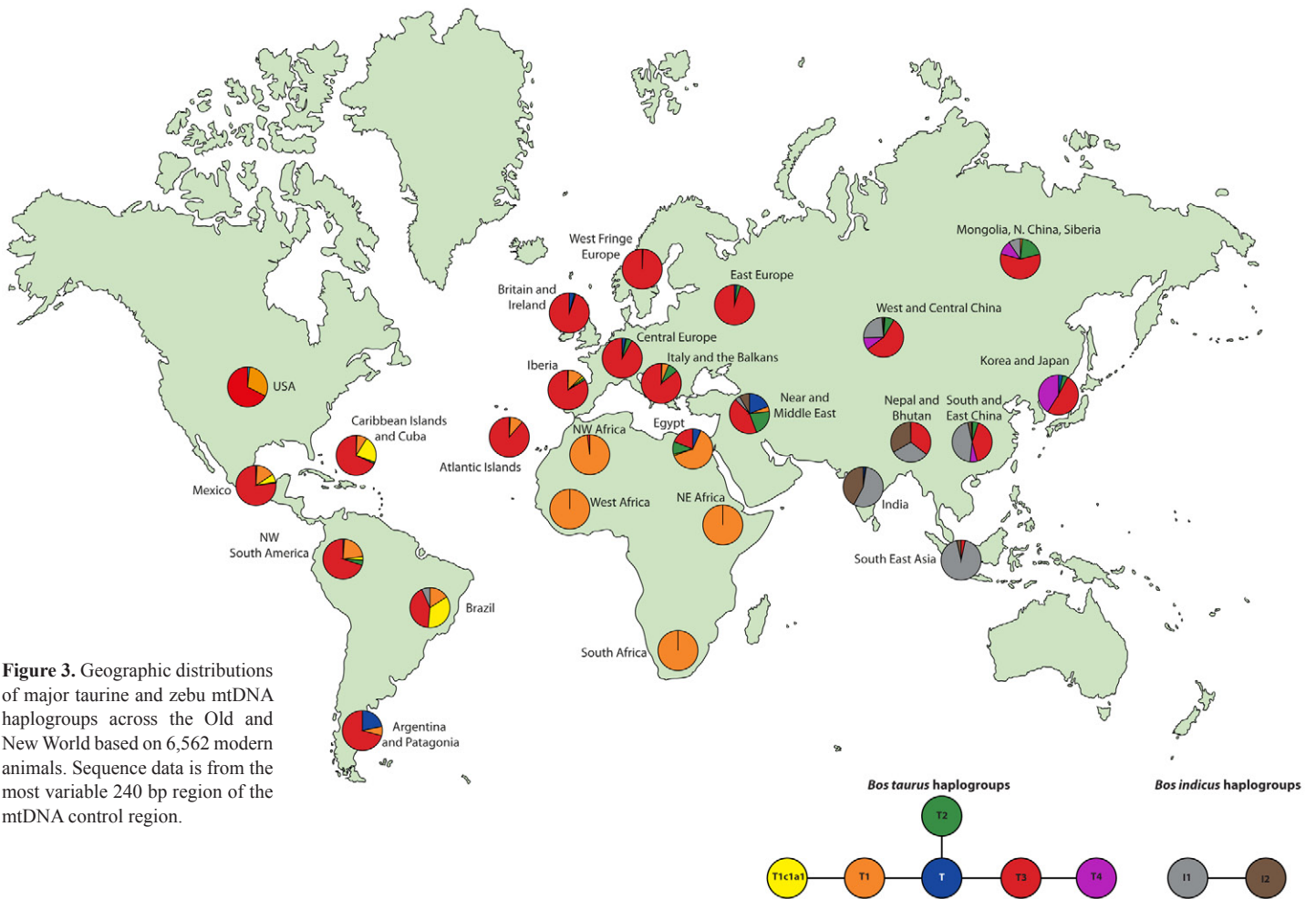


Figure 3. Geographic distributions of major taurine and zebu mtDNA haplogroups across the Old and New World based on 6,562 modern animals. Sequence data is from the most variable 240 bp region of the mtDNA control region.

exclusively within the Indian subcontinent and only sporadically outside this region. Current data support northern India as the center for origin for this haplogroup, which expanded sometime after the expansion of I1. The current distribution of I2 across the Indian subcontinent is most readily explained by later diffusion by trade throughout the region (Baig et al., 2005; Magee et al., 2007; Chen et al., 2010). That modern zebu mtDNA sequence variation comprises two phylogenetically distinct, yet closely related, haplogroups reflects both the limited ancestral mtDNA variation incorporated into zebu domestic pools during the domestication process (and which has survived in modern populations) and the limited variation that has accumulated since this time through mutation.

Nuclear Genetic Data Support the Independent Domestic Origins of *B. taurus* and *B. indicus* and the Mosaicism of African Cattle

A major drawback of mtDNA-based studies is that they focus only on a single segregating locus with a uniquely maternal mode of inheritance and do not, therefore, represent ancestry across the whole genome. For example, the effects of male-mediated gene flow, which also underlies modern animal breeding and selection strategies, cannot be discerned through the analysis of this exclusive matrilineal genetic marker. Therefore, it is

important that the information gleaned from mtDNA is complemented with analyses of autosomal and Y chromosomal polymorphisms.

Analyses of autosomal and Y chromosomal genetic markers (Box 3) also show the marked divergence between the two domestic taxa, as discerned by mtDNA analysis. In particular, these nuclear studies have revealed that alternate collections of alleles from several nuclear loci display high frequencies in either modern taurine and zebu populations, respectively, providing further evidence of separate domestic origins of the two major domestic types and the pronounced time depth in their genetic separation (MacHugh et al., 1997; Hanotte et al., 2002).

Nuclear markers have also been instrumental in teasing out the historical migratory routes of cattle from their centers of origin across the Old World as a consequence of early pastoralist diasporas. This is particularly evident in Africa, where analysis of nuclear genetic markers has shown that modern African zebu populations possess autosomal genomes more similar to Indian *B. indicus* despite all sampled modern African cattle having mtDNA sequences that are clearly *B. taurus* in origin (MacHugh et al., 1997; Hanotte et al., 2002; Bovine HapMap Consortium, 2009; Lenstra et al., 2014). This finding is further supported by the use of autosomal and Y chromosomal zebu-specific markers (that is, markers that are present at high frequencies in Indian *B. indicus* populations compared with *B. taurus* populations), where a cline of east–west zebu-derived alleles running in an east–west direction across the continent and a north–south direction in West Africa is evident (Bradley et al., 1994; MacHugh et al., 1997; Free-

Box 4. Ancient DNA

The study of ancient DNA is a sub-discipline of biomolecular archaeology, which encompasses extraction of any biomolecule (such as DNA, blood proteins and collagen fibers, resins, fats, and oils) from ancient tissues. The analysis of DNA from archaeological and palaeontological samples has the potential to answer questions about the past directly, thus providing a temporal aspect to modern genetic studies. Ancient studies have shown that DNA can survive intact for more than 500,000 years, enabling many evolutionary questions to be addressed over this time period, including biogeography, phylogeny, taxonomy and selective processes at the population level, as well as domestication origins. Ancient DNA can be extracted from a variety of animal remains, ranging in condition and age, the most popular of which are bones, teeth, and hair. The cytoplasmic location and high copy number of mitochondrial chromosomes within each eukaryotic cell (estimated at up to 10^3 molecules per cell) has made mtDNA the primary genetic marker for ancient DNA studies. However, recent advances in genomic technologies have seen an increase in the number of ancient DNA studies that involve partial or whole nuclear genome sequences from archaeological samples (Shapiro and Hofreiter, 2014).

man et al., 2004; Freeman et al., 2006). The apparent phylogenetic disparity between African mtDNA and nuclear markers is presumably attributable to predominantly male-mediated zebu introgression from India into Africa as a result of large-scale Arabic migrations from AD 711 onward. Consequently, the founding taurine genetic substrate of Africa has been eroded and displaced by successive waves of zebu migration from Asia and Arabia into the continent. To date, only the trypanotolerant taurine populations of West Africa (such as the N'Dama population of Guinea) have not been subject to appreciable zebu genetic introgression (Bradley et al., 1994; MacHugh et al., 1997).

Phylogenomic analyses of high-density SNP genotyping data involving tens of thousands of biallelic loci (Box 3) have enabled fine-scaled population genetic analyses previously undetected from mtDNA and STR data. For example, Gautier et al. (2010) showed that unsupervised statistical analysis of pan-genomic SNP data permitted samples to be unambiguously assigned to their population of origin; thus, illustrating the power of high-density SNP data to assess the fine-scale genetic structure of modern breeds. These results reflect the differences in allele frequencies that exist between breeds, including those of similar geography and with shared coancestry. These genetic differences have been generated over the past 200 years due largely to the subsampling of ancestral variation and evolutionary and reproductive isolation.

More importantly, high-density SNP data have enabled detailed high-resolution phylogenomic analysis from which detailed descriptions of cattle domestic history can be elucidated (Decker et al., 2009; Bovine HapMap Consortium, 2009; Gautier et al., 2010; Decker et al., 2014). As with early studies involving mtDNA and STR loci, the data demonstrate the pre-domestic divergence of taurine and zebu cattle, supporting their separate domestic origins (Figure 4). Migrations (and subsequent hybridization) of animals from domestic centers of origin resulted in the appreciable levels of zebu and taurine admixture present in the morphologically taurine and zebu populations of Asia, respectively. The data further

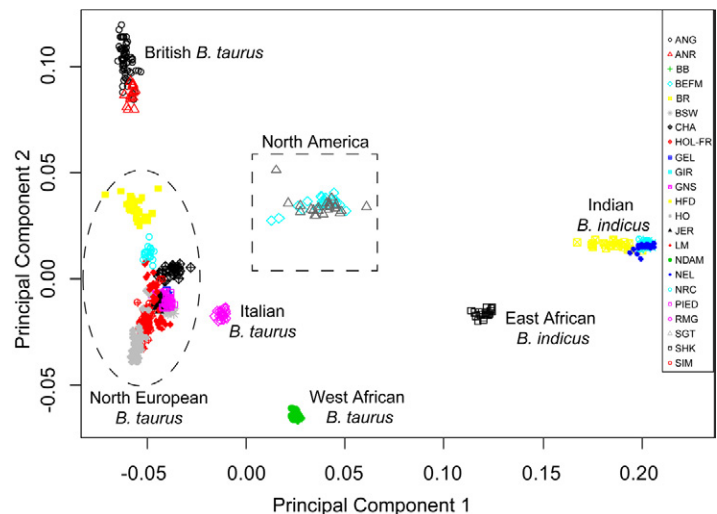


Figure 4. Principal component (PC) analysis of modern taurine and zebu populations based on 44,000 autosomal SNPs. Each symbol represents a single animal. Taurine breeds are separated from zebu breeds along PC1 with admixed breeds from North America and Africa occupying intermediate positions. The Romagnola breed of Italy is separated from the main European taurine group along PC1, suggesting zebu admixture in this population. PC2 partitions breeds within the taurine lineage, with British taurine and West African taurine populations at the extremities of PC2; continental European taurine populations occupy intermediate positions along PC2. Breed codes: Europe *B. taurus* (ANG, Angus; ANR, Angus Red; BB, Belgian Blue; BSW, Brown Swiss; CHA, Charolais; HOL-FR, Holstein-Friesian; GEL, Gelbvieh; GNS, Guernsey; HFD, Hereford; HOL, Holstein; JER, Jersey; LM, Limousin; NRC, Norwegian Red; PIED, Piedmontese; RMG, Romagnola; SIM, Simmental); African *B. taurus* (NDAM, Guinean N'Dama); African *B. indicus* (SHK, Shoko); America (BEFM, Beefmaster; SGT, Santa Gertrudis); *B. indicus* (BR, Brahman; GIR, Gir; NEL, Nelore).

confirm the zebu ancestry of African cattle, ranging from 0–20% in West African taurine populations and increasing to 23–74% in southern, central, and eastern African hybrid populations (Figure 4; Decker et al., 2009, 2014). Zebu ancestry has also been detected in modern Italian populations (but not in northern and central European and Iberian populations), which may reflect the Roman importation of Near Eastern cattle (in which zebu admixture had already occurred) to South Europe (Decker et al., 2014). Furthermore, partial African taurine ancestry of Iberian and Italian cattle was confirmed and is presumably a legacy of historical interactions across the Mediterranean.

Despite African taurine cattle being more divergent from European and Asian taurine populations (Figure 4), pan-genomic SNP analyses show that taurine cattle from all three continents shared a single recent common ancestor, supporting a single Near Eastern domestic origin for *B. taurus* (Decker et al., 2014). Migrations of taurine domesticates from this center of origin was followed by admixture with Asian and African aurochs with possible minor captures of indigenous aurochs (Decker et al., 2009, 2014). It is also postulated that the Near Eastern-derived mtDNA haplotypes of early African domesticates was replaced with the divergent T1 haplogroup as a result of the hybridization between migrating domestic populations and resident wild African populations (Decker et al., 2014). Purifying natural selection, in which certain wild mtDNA haplogroups conferred an advantage in African environments, may have also accelerated replacement of Near Eastern-derived haplotypes in Africa (Soares et al., 2013). Analysis also supported the introgression of southern European

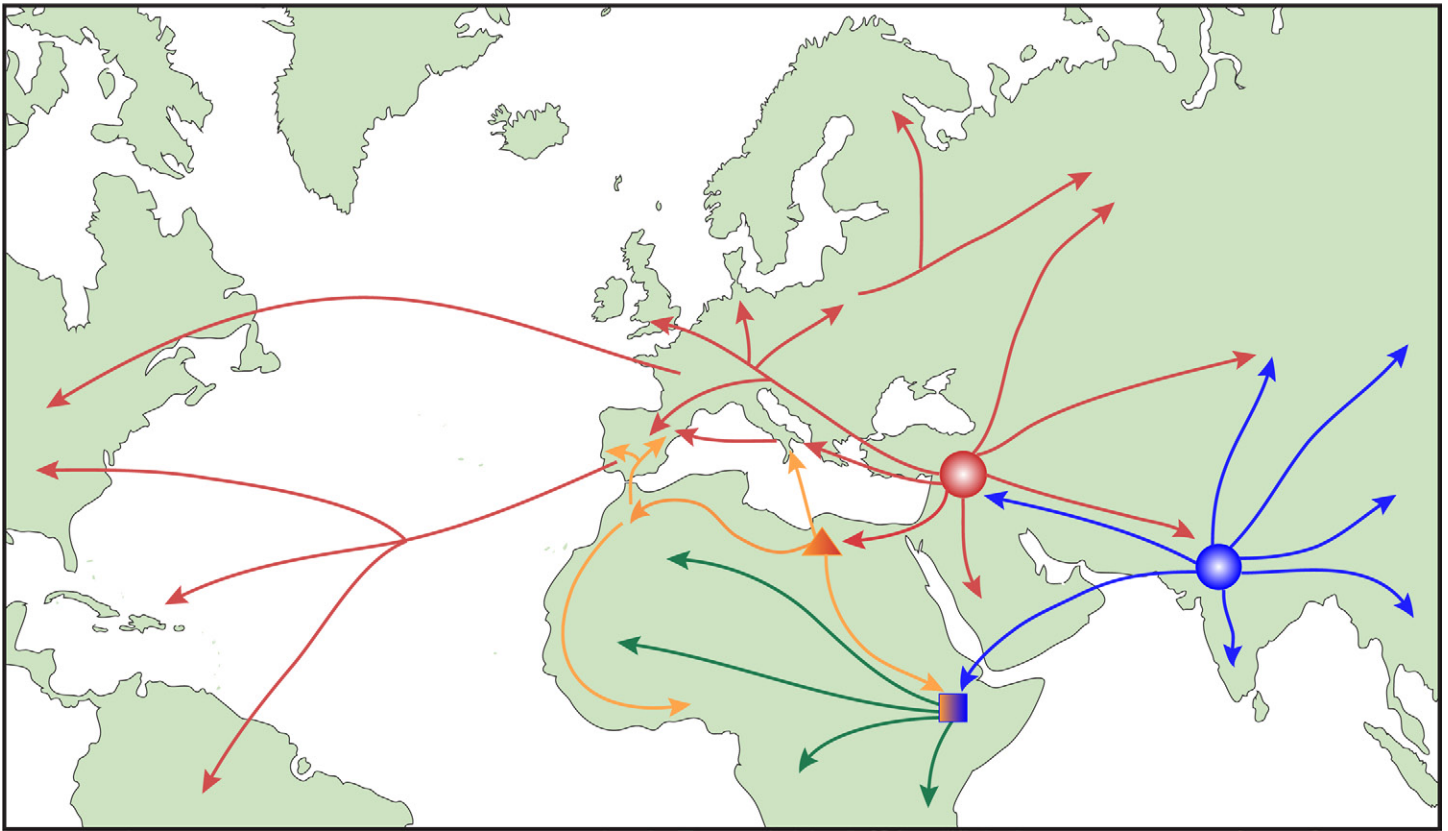


Figure 5. The migratory routes of domestic cattle from their centers of origin based on high-resolution phylogenomic analysis and interpretation of high-density autosomal SNP data. Centers of taurine and zebu domestication are shown as red and blue circles, respectively. Movements of Near Eastern taurine animals (red line) into Europe via Danubian and Mediterranean routes *circa* 9,000 to 8,000 YBP gave rise to the founding *B. taurus* of the continent. Near Eastern taurine movements into Africa (*circa* 8,000 YBP) were followed by admixture with indigenous African aurochs populations (shaded triangle; the movements of these animals are shown as an orange line) resulting in the distinct genomic structure of modern African taurine populations. The African taurine ancestry of Iberian and Italian populations support the movement of African domesticates into Southern Europe across the Mediterranean Sea, presumably due, in part, to the Moorish occupation of Iberia. Pastoralist migrations from the two major centers of domestication (zebu migrations are shown as a blue line) across the Iranian Plateau *circa* 4,000 YBP led to admixture between zebu and taurine populations, resulting in the mixed ancestry of modern Near Eastern taurine and North Indian zebu populations. Zebu cattle entered Africa ~1,300 years ago where they hybridized with the founding taurine populations of the continent (shaded square), giving rise to the admixed populations of east, central, and southern Africa (green lines). Iberian cattle were imported to the Americas during the 15th and 16th centuries AD; the African genetic influence observed in modern Creole populations originated in these Iberian imports. British and European cattle entered North America during the 17th and 18th centuries AD. The zebu ancestry of American cattle is a legacy of introductions from India during the 19th century AD.

cattle into Far East Asian populations, resulting in the introduction of European taurine and African taurine and zebu genetic contributions (a feature of southern European populations) to the region (Decker et al., 2009, 2014). The current model for the recent evolution and history of domestic cattle, which is based on the analysis and interpretation of high-density SNP data, is presented in Figure 5.

New World Cattle Display Genetic Ancestry from Multiple Independent Locations

The European discovery and conquest of the Americas prompted the movement of people and cattle to the New World. Domestic cattle did not exist on the American continent until AD 1493, when Christopher Columbus landed a small number of Spanish cattle on the Caribbean Island of Hispanola. Cattle were continually imported to the region over the following 50 years as agro-pastoralism intensified (Wilkins, 1984), and the majority of the cattle transported to the Americas during this time were of Andalusian (southwest Spain) origin. However, some authors contend

that cattle from the Canary Islands, which had been populated with breeds from northwest Spain, were also introduced (Primo, 1992). By the mid-16th century AD, cattle of Spanish ancestry populated most Caribbean islands and had reached Central and South America; these animals are the ancestors of the modern *Criollo* breeds. The 16th century AD also witnessed the importation of Portuguese cattle to Brazil, imported either directly from Portugal or from the Cape Verde Islands (de Alba, 1978). In addition, trade links between northern Europe and North America, as well as Australia, established as part of the European colonization of North America in the 17th century, led to the introduction of several European populations, which were subsequently crossbred to generate the hybrid breeds that predominate in North America today.

Between the 16th and 18th centuries AD, African *B. taurus* cattle were introduced to the Americas, possibly as a consequence of the Atlantic slave trade and secondary economic activity. It is believed that these cattle were shipped directly from the French, Portuguese, and Spanish colonies located on the Guinean Gulf coast (Maillard et al., 1993; Maillard and Maillard, 1998). Purchases of West African cattle by the later-established

French settlers of Martinique and Guadeloupe during the 17th and 18th century AD have also been reported (Maillard and Maillard, 1998). Towards the beginning of the 19th century AD, importations of zebu cattle became widespread throughout the Americas. Zebu animals were initially imported from Senegal and were employed to supply the traction necessary for crop cultivation (Rouse, 1973). During the 20th century AD, African zebu animals were gradually replaced by Indian zebu animals, which exhibit increased durability in tropical environments. Crossbreeding of zebu cattle with native Creole animals led to marked rises in cattle productivity in tropical regions, and consequently, large-scale importations of Indian zebu cattle became widespread throughout the Americas during this time (Wilkins, 1984).

Mitochondrial DNA sequence analyses of modern Caribbean and Central and South American cattle reflect the multifaceted historical importations of Eurasian cattle to the Americas. These studies have revealed the predominance of European T3 haplogroup sequences in these populations with varying, yet considerable, frequencies of sequences belonging to the T1 haplogroup. Of note, the derivative T1 haplotype, T1c1a1, which is only sporadically encountered in Iberian populations, has been detected at frequencies of 31% in the populations of the Caribbean Lesser Antilles islands and 50% in modern Brazilian Criollo populations (Figure 2; Magee

et al., 2002; Carvajal-Carmona et al., 2003; Mirol et al., 2003; Miretti et al., 2004; Ginja et al., 2010).

The numerical predominance of a peripheral T1 derivative haplotype within these populations suggests a founder effect, in which a restricted subset of the African-derived variation within the Iberian parental population has been retained in modern New World populations. However, this finding does not preclude the direct importations of African cattle to the region. Indeed, genetic studies have revealed the presence of zebu-specific STR alleles *sans* zebu mtDNA sequences, a feature of African zebu populations, within modern Caribbean populations, thus supporting West African zebu cattle importations to the Americas (Magee et al., 2002; Liron et al., 2006). This is further supported by phylogenetic analyses of New World cattle involving STR markers, which demonstrate that African zebu and taurine cattle contributed significant genetic components of modern American cattle (Delgado et al., 2012; Martinez et al., 2012). Recent analysis of high-density SNP data confirm the genetic affinity between New World and Iberian cattle and has revealed levels of African admixture in modern American breeds that is higher than previously estimated from historical records and genetic analyses based on STR and mtDNA data; these results suggest that the African ancestry observed in modern American populations is, in part, of direct African origin (Gautier and Naves, 2011). However, more recent SNP-based studies of American cattle genetic diversity suggests that this African genetic influence was present in Iberian cattle before the introduction of cattle to the New World; these findings support an Iberian origin for the African ancestry observed in modern New World populations (McTavish et al., 2013; Decker et al., 2014).

More recent Indian zebu genetic influence in New World populations is supported by several lines of evidence including mtDNA (Magee et al., 2002; Paneto et al., 2008), Y chromosome microsatellites (Giovambattista et al., 2000), and autosomal STR and SNP genetic markers (Liron et al., 2006; Egitto et al., 2007; Achilli et al., 2009; McTavish et al., 2013). Interestingly, McTavish et al. (2013) suggest that the zebu ancestry observed in New World cattle may have originated, in part, from the founding Iberian cattle of the Americas; however, the lack of zebu ancestry in Iberian cattle has led Decker and colleagues (2014) to refute this and propose that zebu introgression in the Americas occurred after the arrival of Iberian cattle in the New World.

The Promise of Ancient DNA for Studies of Cattle Domestication

Due to the high copy number of mitochondrial genomes in the cell compared with the nuclear genome, ancient DNA research in cattle has been concentrated primarily on the analysis of mtDNA variation (Box 4; MacHugh et al., 1999; Troy et al., 2001; Bollongino and Burger, 2007; Edwards et al., 2007b; Stock et al., 2009). Studies have been successful in addressing some broad questions, such as the nature of domestication and the relationship of domestic cattle to the European wild aurochs. Mitochondrial DNA sequences from aurochs skeletal remains sampled from central and northern Europe (including Britain) belong largely to a single haplogroup, P, which is phylogenetically differentiated from modern European *B. taurus* animals (Figures 1B and 1C; Troy et al., 2001; Edwards et al., 2007b). In addition to haplogroup P, another haplotype, E, was detected in a single specimen from the Early Neolithic site of Eilsleben in Germany (Edwards et al., 2007b).



The absence of European aurochs mtDNA sequences in modern European *B. taurus* animals supports the hypothesis that autochthonous aurochs were not domesticated within Europe, but instead that domestic cattle were derived primarily from populations domesticated in the Near East (Troy et al., 2001; Edwards et al., 2007b). However, localized aurochs domestication within Europe cannot be completely discounted; Achilli et al. (2009) identified two novel haplogroups (Q and R) in modern Italian cattle that are considerably different to those previously described in modern *B. taurus* populations. Also, a single European *B. primigenius*-defining haplogroup P sequence obtained from a modern animal has been deposited in the GenBank sequence repository (Achilli et al., 2008) while another haplotype (haplotype C), has been identified in an archaeological assemblage from early Holocene China (Figure 1C; Zhang et al., 2013). Collectively, these data support minor captures of wild matriline during the colonization of Europe. In addition to mtDNA, ancient DNA studies using nuclear markers have aimed to address the long-standing question of the localized domestication of European aurochs or male-mediated introgression from the wild into domestic herds; however, results from these studies have remained equivocal (Götherström et al., 2005; Bollongino et al., 2006; Svensson and Götherström, 2008).

Despite the availability of a large set of modern data, the information content of cattle mitochondrial haplotypes limits their use for ancient DNA studies; while valuable for global phylogeography, current western European cattle populations display little mitochondrial population genetic structure (Lenstra et al., 2014). On the other hand, nuclear DNA data have been shown to provide a breed-level genetic resolution within modern cattle and thus a more nuanced picture of genetic variation across Europe (Bovine HapMap Consortium, 2009; Gautier et al., 2010; Edwards et al., 2011). Therefore, where possible, extension of nuclear DNA-based studies from extant populations to archaeological specimens will greatly help our understanding of ancient population affinities and local domestication events.

New Genomic Technologies and the Future of Cattle Domestication Studies

The availability of a well-annotated taurine bovine genome sequence (Bovine Genome Sequencing and Analysis Consortium et al., 2009), with associated advances in high-throughput genomic technologies, have revolutionized genomic studies and immediately suggests powerful new approaches to better understand cattle domestication. Furthermore, high-throughput resequencing projects using so-called “next-generation” technologies, whereby millions of genomic DNA fragments from an individual are randomly generated, sequenced in parallel and analyzed, have generated whole-genome sequences for several animals, including animals of economic importance (Larkin et al., 2012).

The domestication of cattle offers a highly valuable model to study evolution; since their domestication, cattle have been continuously exposed to both natural and artificial selection, and these processes, coupled with genetic drift, have produced the multitude of breeds, phenotypes, and production characteristics evident today. Phenotypes selected by humans post-domestication include milk and meat production and quality, docility and temperament, fertility, and relatively aesthetic traits (such as coat color and polledness; Zeder et al., 2006). The selection processes influencing these phenotypes have left indelible marks on the genome of modern populations that can be detected via the analysis of data from high-throughput

genotyping and sequencing platforms. For example, selection for beneficial genetic variants that control or contribute to a desired phenotype can result in the loss of sequence variation in the DNA flanking the selected variant and ultimately the fixation in the population of the chromosomal haplotype that harbors the selected variant. Chromosomal haplotypes that show fixation as a result of positive selection are said to be subject to a “selective sweep.” Analysis of selective sweeps can be used to identify regions of the genome that underlie important phenotypes. These traits have been subject to domestication-associated selective processes and, more recently, selection acting during breed formation and improvement, which may uncover the identity of genes and regulatory segments that have large phenotypic effects (Nielsen, 2005).

Evidence for selective sweeps has been detected in several pan-genomic surveys of bovine SNP diversity and resequencing projects. Results from the Bovine HapMap project identified selective sweeps centered around the myostatin (*MSTN*) and the ATP binding cassette sub family G member 2 (*ABCG2*) genes, which harbor SNPs associated with phenotypic differences in meat and milk production, respectively (Bovine HapMap Consortium, 2009). More recent studies have identified additional genomic regions subject to selective sweeps that encompassed quantitative trait loci and candidate genes associated with beef and dairy production traits, behavioral traits, morphology, fertility, and health (Larkin et al., 2012; Qanbari et al., 2012; Ramey et al., 2013; Rothhammer et al., 2013; Utsunomiya et al., 2013). It is important to note, however, that analysis of selective sweeps is challenging; for example, regions of the genome with significantly reduced sequence variation can also be generated by genetic drift, making true population genomic signals of selection difficult to discern fully.

Perhaps the most exciting application of high-throughput genomic technologies for investigating cattle domestication is through analysis of ancient pre-domestic whole genomes. In particular, NGS is revolutionizing paleogenetics, and considerable genomic DNA sequence information has already been reported from extinct mammals, including hominids (Miller et al., 2008, 2009; Green et al., 2010; Orlando et al., 2013). Until recently, reconstruction of a complete aurochs genome seemed unimaginable, but the first complete aurochs mtDNA genome sequence was published in 2010 (Edwards et al., 2010) using NGS technologies (in combination with conventional DNA sequencing). This sample was from an archaeologically verified British *B. p. primigenius* bone, radiocarbon-dated to 6,700 cal. YBP, more than 1,000 years before the arrival of domestic *B. taurus* to Britain (Rowley-Conwy, 2011). In addition to facilitating reconstruction of the mtDNA genome, this analysis also generated a large number of DNA sequences that mapped to the nuclear *B. taurus* genome, a finding that bodes well for generation of a complete *B. primigenius* genome sequence. Analysis of the genome sequence of the aurochs would provide a reference template for comparison of modern bovine genomes, providing the opportunity to more fully investigate the selective processes that have shaped modern cattle genomes since domestication more than 10,000 years ago. In this context, comparisons of both domestic and non-domestic genomes can answer fundamental questions about the evolutionary process and how it shapes biological variation in animal populations.

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Literature Cited

- Achilli, A., S. Bonfiglio, A. Olivieri, A. Malusà, M. Pala, B. Hooshiar Kashani et al. 2009. The multifaceted origin of taurine cattle reflected by the mitochondrial genome. *PLoS ONE* 4:E5753.
- Achilli, A., A. Olivieri, M. Pellecchia, C. Ubaldi, L. Colli, N. Al-Zahery, M. Accetturo et al. 2008. Mitochondrial genomes of extinct aurochs survive in domestic cattle. *Curr. Biol.* 18:R157–R158.
- Allchin, B., and R. Allchin. 1968. The birth of Indian civilization: India and Pakistan before 500 B.C. Penguin Books, New York.
- Allchin, F.R., and B. Allchin. 1974. The relationship of Neolithic and later settled communities with those of Late Stone Age hunters and gatherers. In: D.D. Kosambi and R.S. Sharma, editors, *Indian society: Historical probings, in memory of D. D. Kosambi*. People's Publishing House, New Delhi. p. 45–66.
- Anderung, C., A. Bouwman, P. Persson, J.M. Carretero, A.I. Ortega, R. Elburg et al. 2005. Prehistoric contacts over the Straits of Gibraltar indicated by genetic analysis of Iberian Bronze Age cattle. *Proc. Natl. Acad. Sci. USA* 102:8431–8435.
- Baig, M., A. Beja-Pereira, R. Mohammad, K. Kulkarni, S. Farah, and G. Luikart. 2005. Phylogeography and origin of Indian domestic cattle. *Curr. Sci.* 89:38–40.
- Baker, C.M., and C. Manwell. 1980. Chemical classification of cattle: 1. Breed groups. *Anim. Blood Groups Biochem. Genet.* 11:127–150.
- Bandelt, H.J., P. Forster, B.C. Sykes, and M.B. Richards. 1995. Mitochondrial portraits of human populations using median networks. *Genetics* 141:743–753.
- Bar-Yosef, O. 2011. Climatic fluctuations and early farming in West and East Asia. *Curr. Anthropol.* 52:S175–S193.
- Beja-Pereira, A., D. Caramelli, C. Lalueza-Fox, C. Vernesi, N. Ferrand, A. Casoli, F. Goyache et al. 2006. The origin of European cattle: Evidence from modern and ancient DNA. *Proc. Natl. Acad. Sci. USA* 103:8113–8118.
- Bökönyi, S. 1976. Development of early stock rearing in the Near East. *Nature* 264:19–23.
- Bökönyi, S. 1997. Zebus and Indian wild cattle. *Anthropozoologica* 25–26:647–654.
- Bollongino, R., and J. Burger. 2007. Neolithic cattle domestication as seen from ancient DNA. *Proc. Br. Acad.* 144:165–187.
- Bollongino, R., C.J. Edwards, K.W. Alt, J. Burger, and D.G. Bradley. 2006. Early history of European domestic cattle as revealed by ancient DNA. *Biol. Lett.* 2:155–159.
- Bovine Genome Sequencing and Analysis Consortium, C.G. Elsik, R.L. Tellam, and K.C. Worley. 2009. The genome sequence of taurine cattle: A window to ruminant biology and evolution. *Science* 324:522–528.
- Bovine HapMap Consortium. 2009. Genome-wide survey of SNP variation uncovers the genetic structure of cattle breeds. *Science* 324:528–532.
- Bradley, D.G., D.E. MacHugh, R.T. Loftus, R.S. Sow, C.H. Hoste, and E.P. Cunningham. 1994. Zebu-aurine variation in Y chromosomal DNA: A sensitive assay for genetic introgression in west African trypanotolerant cattle populations. *Anim. Genet.* 25:7–12.
- Bradley, D.G., and D.A. Magee. 2006. Genetics and the origins of domestic cattle. In: M.A. Zeder, D.G. Bradley, E. Emshwiller, and B.D. Smith, editors, *Documenting domestication: New genetic and archaeological paradigms*. University of California Press, Berkeley. p. 317–328.
- Brown, A. 2007. Dating the onset of cereal cultivation in Britain and Ireland: The evidence from charred cereal grains. *Antiquity* 81:1042–1052.
- Budiński, S. 1994. The covenant of the wild: Why animals chose domestication. Yale University Press, New Haven, CT.
- Carvajal-Carmona, L.G., et al. 2003. Abundant mtDNA diversity and ancestral admixture in Colombian criollo cattle (*Bos taurus*). *Genetics* 165:1457–1463.
- Chen, S., N. Bermudez, M. Olivera-Angel, L. Estrada, J. Ossa, G. Bedoya, and A. Ruiz-Linares. 2010. Zebu cattle are an exclusive legacy of the South Asia Neolithic. *Mol. Biol. Evol.* 27:1–6.
- Clason, A.T. 1978. Late Bronze-Age- Iron-Age zebu cattle in Jordan. *J. Archaeol. Sci.* 5:91–93.
- Clutton-Brock, J. 2012. *Animals as domesticates: A world view through history*. Michigan State Univ. Press, East Lansing.
- Cymbron, T., R.T. Loftus, M.I. Malheiro, and D.G. Bradley. 1999. Mitochondrial sequence variation suggests an African influence in Portuguese cattle. *Proc. R. Soc. B* 266:597–603.
- de Alba, J. 1978. *Progress in the selection of the Latin American dairy Criollo*. FAO, Rome, Italy.
- Decker, J.E., S.D. McKay, M.M. Rolf, J.W. Kim, A.M. Alcalá, T.S. Sonstegard et al. 2014. Worldwide patterns of ancestry, divergence, and admixture in domesticated cattle. *PLoS Genet.* 10:E1004254.
- Decker, J.E., J.C. Pires, G.C. Conant, S.D. McKay, M.P. Heaton, K. Chen, A. Cooper et al. 2009. Resolving the evolution of extant and extinct ruminants with high-throughput phylogenomics. *Proc. Natl. Acad. Sci. USA* 106:18644–18649.
- Delgado, J.V., A.M. Martínez, A. Acosta, L.A. Alvarez, E. Armstrong, E. Camacho, J. Cañón et al. 2012. Genetic characterization of Latin-American Creole cattle using microsatellite markers. *Anim. Genet.* 43:2–10.
- Diamond, J. 2002. Evolution, consequences and future of plant and animal domestication. *Nature* 418:700–707.
- Dolukhanov, P., A. Shukurov, D. Gronenborn, D. Sokoloff, V. Timofeev, and G. Zaitseva. 2005. The chronology of neolithic dispersal in Central and Eastern Europe. *J. Archaeol. Sci.* 32:1441–1458.
- Edwards, C.J., J.F. Baird, and D.E. MacHugh. 2007a. Taurine and zebu admixture in Near Eastern cattle: A comparison of mitochondrial, autosomal and Y-chromosomal data. *Anim. Genet.* 38:520–524.
- Edwards, C.J., R. Bollongino, A. Scheu, A. Chamberlain, A. Tresset, J.D. Vigne, and J.F. Baird et al. 2007b. Mitochondrial DNA analysis shows a Near Eastern Neolithic origin for domestic cattle and no indication of domestication of European aurochs. *Proc. Biol. Sci.* 274:1377–1385.
- Edwards, C.J., C. Ginja, J. Kantanen, L. Pérez-Pardal, A. Tresset, F. Stock, et al. 2011. Dual origins of dairy cattle farming—evidence from a comprehensive survey of European Y-chromosomal variation. *PLoS ONE* 6:E15922.
- Edwards, C.J., D.A. Magee, S.D.E. Park, P.A. McGettigan, A.J. Lohan, A. Murphy, E.K. Finlay et al. 2010. A complete mitochondrial genome sequence from a mesolithic wild aurochs (*Bos primigenius*). *PLoS ONE* 5:E9255.
- Egito, A.A., S.R. Paiva, M.d.S.M. Albuquerque, A.S. Mariante, L.D. Almeida, S.R. Castro, and D. Grattapaglia. 2007. Microsatellite based genetic diversity and relationships among ten Creole and commercial cattle breeds raised in Brazil. *BMC Genet.* 8:83.
- Epstein, H., and I.L. Mason. 1984. *Evolution of domesticated animals*. Longman, London.
- FAO. 2012. *FAO Statistical Yearbook 2012: Europe and Central Asia food and agriculture*. Food and Agriculture Organization of the United Nations, Rome.
- Freeman, A.R., C.J. Hoggart, O. Hanotte, and D.G. Bradley. 2006. Assessing the relative ages of admixture in the bovine hybrid zones of Africa and the Near East using X chromosome haplotype mosaicism. *Genetics* 173:1503–1510.
- Freeman, A.R., C.M. Meghen, D.E. MacHugh, R.T. Loftus, M.D. Achukwi, A. Bado, B. Sauveroche, and D.G. Bradley. 2004. Admixture and diversity in West African cattle populations. *Mol. Ecol.* 13:3477–3487.
- Gautier, M., D. Laloe, and K. Moazami-Goudarzi. 2010. Insights into the Genetic History of French Cattle from Dense SNP Data on 47 Worldwide Breeds. *PLoS ONE* 5(9):e13038.
- Gautier, M., and M. Naves. 2011. Footprints of selection in the ancestral admixture of a New World Creole cattle breed. *Mol. Ecol.* 20:3128–3143.
- Ginja, C., M.C. Penedo, L. Melucci, J. Quiroz, O.R. Martínez López, M.A. Reviddatti, A. Martínez-Martínez, J.V. Delgado, and L.T. Gama. 2010. Origins and genetic diversity of New World Creole cattle: Inferences from mitochondrial and Y chromosome polymorphisms. *Anim. Genet.* 41:128–141.
- Giovambattista, G., M.V. Ripoli, J.C. De Luca, P.M. Mirol, J.P. Lirón, and F.N. Dulout. 2000. Male-mediated introgression of *Bos indicus* genes into Argentine and Bolivian Creole cattle breeds. *Anim. Genet.* 31:302–305.
- Götherström, A., C. Anderung, L. Hellborg, R. Elburg, C. Smith, D.G. Bradley, and H. Ellegren. 2005. Cattle domestication in the Near East was followed by hybridization with aurochs bulls in Europe. *Proc. R. Soc. B* 272: 2345–2350.
- Green, R.E., J. Krause, A.W. Briggs, T. Maricic, U. Stenzel, M. Kircher et al. 2010. A draft sequence of the Neandertal genome. *Science* 328:710–722.
- Grigson, C. 1978. The craniology and relationships of four species of *Bos*. 4. The relationship between *Bos primigenius* Boj. and *Bos taurus* L. and its implications for the phylogeny of the domestic breeds. *J. Archaeol. Sci.* 5:123–152.
- Grigson, C. 1980. The craniology and relationships of four species of *Bos*. 5. *Bos indicus* L. *J. Archaeol. Sci.* 7:3–32.
- Grigson, C. 2000. *Bos africanus* (Brehm)? Notes on the archaeozoology of the native cattle of Africa. In: R.M. Blench and K.C. MacDonald, editors, *The origins*

- and development of African livestock: Archaeology, genetics, linguistics and ethnography. UCL, London. p. 38–60.
- Guintard, C. 1999. On the size of the ure-ox or aurochs (*Bos primigenius Bojanus*, 1827). In: G.-C. Weniger, editor, Archäologie und Biologie des Aurochsens. Neanderthal Museum, Mettmann. p. 7–21.
- Hanotte, O., D.G. Bradley, J.W. Ochieng, Y. Verjee, E.W. Hill, and J.E. Rege. 2002. African pastoralism: Genetic imprints of origins and migrations. *Science* 296:336–339.
- Helmer, D., L. Gourichon, H. Monchot, J. Peters, and M. Saña Seguí. 2005. Identifying early domestic cattle from Pre-Pottery Neolithic sites on the Middle Euphrates using sexual dimorphism. In: J.-D. Vigne, J. Peters, and D. Helmer, editors, The first steps of animal domestication: New archaeozoological approaches. Oxbow, Oxford. p. 86–95.
- Ho, S.Y.W., G. Larson, C.J. Edwards, T.H. Heupink, K.E. Lakin, P.W. Holland, and B. Shapiro. 2008. Correlating Bayesian date estimates with climatic events and domestication using a bovine case study. *Biol. Lett.* 4:370–374.
- Hourani, A. 1991. A history of the Arab peoples. Faber, London.
- Kantanen, J., C.J. Edwards, D.G. Bradley, H. Viinalass, S. Thessler, Z. Ivanova et al. 2009. Maternal and paternal genealogy of Eurasian taurine cattle (*Bos taurus*). *Heredity* 103:404–415.
- Kruglyak, L., and D.A. Nickerson. 2001. Variation is the spice of life. *Nat. Genet.* 27:234–236.
- Lai, S.J., Y.P. Liu, Y.X. Liu, X.W. Li, and Y.G. Yao. 2006. Genetic diversity and origin of Chinese cattle revealed by mtDNA D-loop sequence variation. *Mol. Phylogenet. Evol.* 38:146–154.
- Larkin, D.M., H.D. Daetwyler, A.G. Hernandez, C.L. Wright, L.A. Hetrick, L. Boucek, S.L. Bachman et al. 2012. Whole-genome resequencing of two elite sires for the detection of haplotypes under selection in dairy cattle. *Proc. Natl. Acad. Sci. USA* 109:7693–7698.
- Lenstra, J.A., P. Ajmone-Marsan, A. Beja-Pereira, R. Bollongino, D.G. Bradley, L. Colli et al. 2014. Meta-analysis of mitochondrial DNA reveals several population bottlenecks during worldwide migrations of cattle. *Diversity* 6:178–187.
- Liron, J.P., P. Peral-García, and G. Giovambattista. 2006. Genetic characterization of Argentine and Bolivian Creole cattle breeds assessed through microsatellites. *J. Hered.* 97:331–339.
- Loftus, R.T., D.E. MacHugh, D.G. Bradley, P.M. Sharp, and P. Cunningham. 1994a. Evidence for two independent domestications of cattle. *Proc. Natl. Acad. Sci. USA* 91:2757–2761.
- Loftus, R.T., D.E. MacHugh, L.O. Ngere, D.S. Balain, A.M. Badi, D.G. Bradley, and E.P. Cunningham. 1994b. Mitochondrial genetic variation in European, African and Indian cattle populations. *Anim. Genet.* 25:265–271.
- MacHugh, D.E., and D.G. Bradley. 2001. Livestock genetic origins: Goats buck the trend. *Proc. Natl. Acad. Sci. USA* 98:5382–5384.
- MacHugh, D.E., M.D. Shriver, R.T. Loftus, P. Cunningham, and D.G. Bradley. 1997. Microsatellite DNA variation and the evolution, domestication and phylogeography of taurine and zebu cattle (*Bos taurus* and *Bos indicus*). *Genetics* 146:1071–1086.
- MacHugh, D.E., C.S. Troy, F. McCormick, I. Olsaker, E. Eythórsdóttir, and D.G. Bradley. 1999. Early medieval cattle remains from a Scandinavian settlement in Dublin: Genetic analysis and comparison with extant breeds. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 354:99–108.
- Magee, D.A., H. Mannen, and D. Bradley. 2007. Duality in *Bos indicus* mtDNA diversity: Support for geographical complexity in zebu domestication. In: M.D. Petraglia and B. Allchin, editors, The evolution and history of human populations in South Asia: Inter-disciplinary studies in archaeology, biological anthropology, linguistics, and genetics. Springer, Dordrecht. p. 385–391.
- Magee, D.A., C. Meghen, S. Harrison, C.S. Troy, T. Cymbron, C. Gaillard, A. Morrow, J.C. Maillard, and D.G. Bradley. 2002. A partial African ancestry for the Creole cattle populations of the Caribbean. *J. Hered.* 93:429–432.
- Maillard, J.C., S.J. Kemp, M. Naves, C. Palin, C. Demangel, A. Accipe, N. Maillard, and A. Bensaid. 1993. An attempt to correlate cattle breed origins and diseases associated with or transmitted by the tick *Amblyomma variegatum* in the French West Indies. *Rev. Elev. Med. Vet. Pays Trop.* 46:283–290.
- Maillard, J.C., and N. Maillard. 1998. Histoire du peuplement bovin et de l'introduction de la tique *Amblyomma variegatum* dans les Iles Françaises des Antilles: Synthèse bibliographique. *Ethnozootecnie* 61:19–36.
- Mannen, H., S. Tsuji, R.T. Loftus, and D.G. Bradley. 1998. Mitochondrial DNA variation and evolution of Japanese black cattle (*Bos taurus*). *Genetics* 150:1169–1175.
- Marshall, F. 1989. Rethinking the role of *Bos indicus* in Sub-Saharan Africa. *Curr. Anthropol.* 30:235–240.
- Marshall, F., and E. Hildebrand. 2002. Cattle before crops: The beginnings of food production in Africa. *J. World Prehist.* 16:99–143.
- Martinez, A.M., L.T. Gama, J. Cañón, C. Ginja, J.V. Delgado, S. Dunner et al. 2012. Genetic footprints of Iberian cattle in America 500 years after the arrival of Columbus. *PLoS ONE* 7:E49066.
- Matukumalli, L.K., C.T. Lawley, R.D. Schnabel, J.F. Taylor, M.F. Allan, M.P. Heaton et al. 2009. Development and characterization of a high density SNP genotyping assay for cattle. *PLoS ONE* 4:E5350.
- McTavish, E.J., J.E. Decker, R.D. Schnabel, J.F. Taylor, and D.M. Hillis. 2013. New World cattle show ancestry from multiple independent domestication events. *Proc. Natl. Acad. Sci. USA* 110:E1398–E1406.
- Meadow, R.H. 1989. Osteological evidence for the process of animal domestication. In: J. Clutton-Brock, editor, The Walking larder: Patterns of domestication, pastoralism, and predation. Unwin Hyman, London, Boston. p. 80–90.
- Meadow, R.H. 1993. Animal domestication in the Middle East: A revised view from the Eastern Margin. In: G. L. Possehl, editor, Harappan civilization: A recent perspective. American Institute of Indian Studies: Oxford & IBH Publishing Company, New Delhi. p. 295–320.
- Melander, Y. 1959. The mitotic chromosomes of some cavicorn mammals (*Bos-Taurus* L., *Bison-Bonassus* L. and *Ovis-Aries* L.). *Hereditas* 45:649–664.
- Miller, W., D.I. Drautz, J.E. Janecka, A.M. Lesk, A. Ratan, L.P. Tomsho, M. Packard et al. 2009. The mitochondrial genome sequence of the Tasmanian tiger (*Thylacinus cynocephalus*). *Genome Res.* 19:213–220.
- Miller, W., D.I. Drautz, A. Ratan, B. Pusey, J. Qi, A.M. Lesk, L.P. Tomsho et al. 2008. Sequencing the nuclear genome of the extinct woolly mammoth. *Nature* 456:387–390.
- Miretti, M.M., S. Dunner, M. Naves, E.P. Contel, and J.A. Ferro. 2004. Predominant African-derived mtDNA in Caribbean and Brazilian Creole cattle is also found in Spanish cattle (*Bos taurus*). *J. Hered.* 95:450–453.
- Mirol, P.M., G. Giovambattista, J.P. Liron, and F.N. Dulout. 2003. African and European mitochondrial haplotypes in South American Creole cattle. *Heredity* 91:248–254.
- Misra, V.N. 2001. Prehistoric human colonization of India. *J. Biosci.* 26:491–531.
- Naik, S.N. 1978. Origin and domestication of zebu cattle (*Bos indicus*). *J. Hum. Evol.* 7:23.
- Nielsen, R. 2005. Molecular signatures of natural selection. *Annu. Rev. Genet.* 39:197–218.
- Orlando, L., A. Ginolhac, G. Zhang, D. Froese, A. Albrechtsen, M. Stiller, M. Schubert et al. 2013. Recalibrating *Equus* evolution using the genome sequence of an early Middle Pleistocene horse. *Nature* 499:74–78.
- Paneto, J.C., J.B. Ferraz, J.C. Balieiro, J.F. Bittar, M.B. Ferreira, M.B. Leite, G.K. Merighe, F.V. Meirelles et al. 2008. *Bos indicus* or *Bos taurus* mitochondrial DNA- comparison of productive and reproductive breeding values in a Guzerat dairy herd. *Genet. Mol. Res.* 7:592–602.
- Payne, W.J.A. 1991. Domestication: A forward step in civilisation. In: C.G. Hickman, editor, Cattle genetic resources. Elsevier Science Publishers, Amsterdam, New York. p. 51–72.
- Pilgrim, G.E. 1947. The evolution of the buffaloes, oxen, sheep and goats. *J. Linn. Soc. London, Zool.* 41:272–286.
- Primo, A.T. 1992. El ganado bovino ibérico en las Americas 500 años despues. *Archivos de Zootecnia* 41:421–432.
- Qanbari, S., T.M. Strom, G. Haberer, S. Weigend, A.A. Gheyas, F. Turner, D.W. Burt et al. 2012. A high resolution genome-wide scan for significant selective sweeps: An application to pooled sequence data in laying chickens. *PLoS ONE* 7:E49525.
- Ramey, H.R., J.E. Decker, S.D. McKay, M.M. Rolf, R.D. Schnabel, and J.F. Taylor. 2013. Detection of selective sweeps in cattle using genome-wide SNP data. *BMC Genomics* 14:382.
- Reingruber, A., and L. Thissen. 2009. Depending on C-14 data: Chronological frameworks in the Neolithic and Chalcolithic of Southeastern Europe. *Radio-carbon* 51:751–770.

Rosenbaum, H.C., and A. Deindard. 1998. Caution before claim: An overview of microsatellite analysis. In: R. Desalle and B. Schierwater, editors, *Molecular approaches to ecology and evolution*. Birkhauser, Basel. p. 87–106.

Rothhammer, S., D. Seichter, M. Forster, and I. Medugorac. 2013. A genome-wide scan for signatures of differential artificial selection in ten cattle breeds. *BMC Genomics* 14:908.

Rouse, J.E. 1973. *Cattle of North America*. 1st ed. University of Oklahoma Press, Norman.

Rowley-Conwy, P. 2011. Westward ho! The spread of agriculture from Central Europe to the Atlantic. *Curr. Anthropol.* 52:S431–S451.

Shapiro, B., and M. Hofreiter. 2014. A paleogenomic perspective on evolution and gene function: New insights from ancient DNA. *Science* 343:1236573.

Smith, B.D. 1995. *The emergence of agriculture*. Scientific American Library: Distributed by W.H. Freeman, New York.

Soares, P., D. Abrantes, T. Rito, N. Thomson, P. Radivojac, B. Li, V. Macaulay et al. 2013. Evaluating purifying selection in the mitochondrial DNA of various mammalian species. *PLoS ONE* 8:E58993.

Stock, F., C.J. Edwards, R. Bollongino, E.K. Finlay, J. Burger, and D.G. Bradley. 2009. Cytochrome b sequences of ancient cattle and wild ox support phylogenetic complexity in the ancient and modern bovine populations. *Anim. Genet.* 40:694–700.

Stock, F., and D. Gifford-Gonzalez. 2013. Genetics and African cattle domestication. *Afr. Archaeol. Rev.* 30:51–72.

Svensson, E., and A. Gotherstrom. 2008. Temporal fluctuations of Y-chromosomal variation in *Bos taurus*. *Biol. Lett.* 4:752–754.

Tresset, A., and J.D. Vigne. 2007. Substitution of species, techniques and symbols at the Mesolithic/Neolithic transition in Western Europe. In: A.W.R. Whittle and V. Cummings, editors, *Going over: The Mesolithic-Neolithic transition in north-west Europe*. Published for the British Academy by Oxford Univ. Press, Oxford. p. 189–210.

Troy, C.S., D.E. MacHugh, J.F. Bailey, D.A. Magee, R.T. Loftus, P. Cunningham, A.T. Chamberlain, B.C. Sykes, and D.G. Bradley. 2001. Genetic evidence for Near-Eastern origins of European cattle. *Nature* 410:1088–1091.

Uerpman, H.P. 1996. Animal domestication- accident or intention? In: D.R. Harris, editor, *The origins and spread of agriculture and pastoralism in Eurasia*. UCL Press, London. p. 227–237.

Underhill, P.A., and T. Kivisild. 2007. Use of Y chromosome and mitochondrial DNA population structure in tracing human migrations. *Annu. Rev. Genet.* 41:539–564.

Utsunomiya, Y.T., A.M.P. O'Brien, T.S. Sonstegard, C.P. Van Tassell, A. Santana do Carmo, G. Mészáros et al. 2013. Detecting loci under recent positive selection in dairy and beef cattle by combining different genome-wide scan methods. *PLoS ONE* 8:E64280.

Vavilov, N. 1926. *Studies on the origin of cultivated plants*. Institut Botanique Appliqué et d'Amélioration des Plantes, Leningrad, Russia.

Waterbolck, H.T. 1968. Food production in Prehistoric Europe. *Science* 162:1093.

Wendorf, F., and R. Schild. 1994. Are the early Holocene cattle in the Eastern Saharan domestic or wild? *Evol. Anthropol.* 3:118–128.

Wendorf, F., and R. Schild. 2001. *Holocene settlement of the Egyptian Sahara*. Vol. 1. The archaeology of Nabta Playa. Kluwer Academic, New York.

Wilkins, J.V. 1984. *Criollo cattle of the Americas*. FAO, Rome, Italy.

Zeder, M.A., E. Emshwiller, B.D. Smith, and D.G. Bradley. 2006. Documenting domestication: The intersection of genetics and archaeology. *Trends Genet.* 22:139–155.

Zeuner, A.F.E. 1963. *A history of domesticated animals*. Hutchinson & Co., London.

Zhang, H., J.L.A. Pajmans, F. Chang, X. Wu, G. Chen, C. Lei, X. Yang et al. 2013. Morphological and genetic evidence for early Holocene cattle management in northeastern China. *Nat. Commun.* 4:2755.

Zilhao, J. 2001. Radiocarbon evidence for maritime pioneer colonization at the origins of farming in west Mediterranean Europe. *Proc. Natl. Acad. Sci. USA* 98:14180–14185.

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