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Piperolein B, isopiperolein B and piperamide C9:1(8E): total synthesis and cytotoxicities†

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Total syntheses of the reported structures of piperolein B, isopiperolein B and piperamide C9:1(8E) have been achieved. The analytical data reported for piperolein B and piperamide C9:1(8E) match the synthetic values, however those for isopiperolein B do not. The cytotoxicities of these three structurally similar compounds against cancer cell lines of different tissue origins were evaluated and the results indicated that these compounds show differential effects on cancer cell viability.

Introduction

Studies of plants of the genus *Piper* have led to the isolation of 277 different amide alkaloids to date.¹ Some species of *Piper* plants are widely used in folk medicine and the isolated amides have been shown to possess a wide range of biological activities. Amongst these, piperolein B, **1** which can be isolated from black pepper, *Piper nigrum*, has been reported to possess larvicidal,² hepatoprotective³ and enzymatic inhibition activity (Fig. 1).⁴ It has also been shown to be a TRP agonist.⁵ Piperamide C9:1(8E), **2** was also tested for larvicidal activity and was found to be six times more toxic than piperolein B, **1**. More recently, isopiperolein B, **3** was isolated.⁶ This was later shown to have antibacterial activity⁷ and cytotoxic activity towards the human cervical carcinoma cell line HeLa.⁸

To date there have been few total syntheses of members of this family of amide alkaloids. However, Strunz and co-workers have synthesised over a dozen of these compounds using a multi-step aldol condensation–Grob-type fragmentation strategy,⁹ whilst several dimeric compounds have been prepared through cycloaddition processes.¹⁰ We were interested in developing a short synthetic route into piperolein B, **1** and the closely related molecules **2** and **3** and investigating their anti-cancer properties.

Results and discussion

A simple synthetic strategy was conceived for the preparation of these compounds (Scheme 1). Amidation of either 8-nonoic acid or 9-decenoic acid with piperidine or pyrrolidine provided the amides **6**, **7** and **8** in good yields. Subsequent Ru-catalyzed cross metathesis with styrene derivative **9** delivered the three natural products **1**, **2** and **3** in moderate yields. These yields of isolated compounds were lower than expected due to difficulties in separating the natural products from the dimer of styrene **9**.

The analytical data for piperolein B, **1** and piperamide C9:1(8E), **2** matched the literature values. However, the data for amide **3** did not match the literature values for isopiperolein B (Table 1). Moreover, the literature values are inconsistent with a pyrrolidine amide and more closely resemble the data for a piperidine amide. This difference can clearly be seen in the aliphatic region of the ¹H NMR

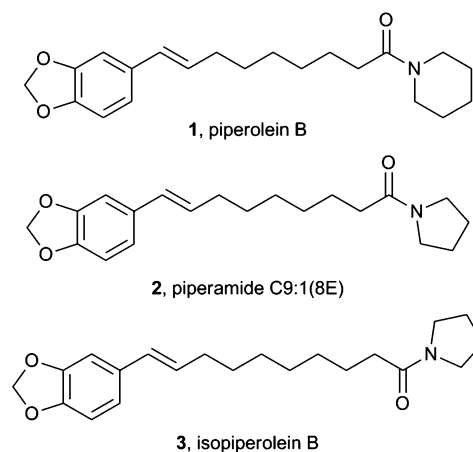


Fig. 1 Reported structures for three *Piper* alkaloids.

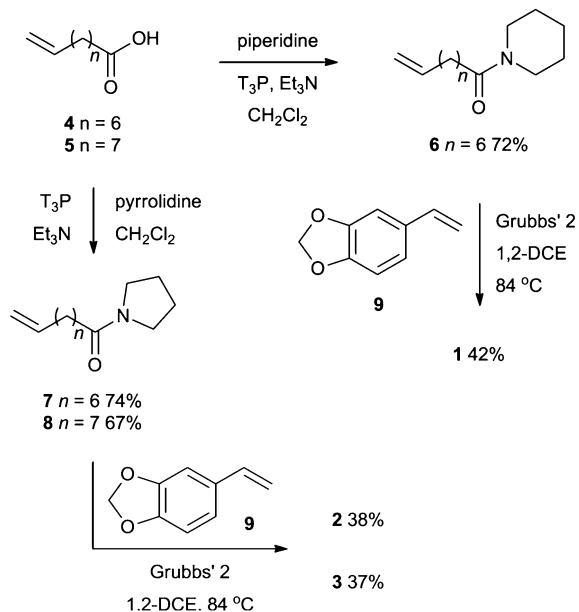
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† Electronic supplementary information (ESI) available: ¹H and ¹³C NMR spectra for compounds **1**, **2**, **3**, **6**, **7** and **8**. See DOI: 10.1039/c3ra42060d

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Scheme 1 Synthesis of alkaloids **1**, **2** and **3**.

spectra of amides **1**, **2** and **3** (Fig. 2). The reported data for isopiperolein B is more like **1** than **3**. It remains unclear as to what the actual structure of this alkaloid is.

As the potential anti-proliferative effects of these compounds on carcinoma cells remain virtually unexplored, we were interested in comparing and contrasting the biological effects of these three structurally very similar amides on cancer cells. Importantly, unlike the majority of previously published studies that routinely involve testing compounds on single cell lines, which in most cases are not closely representative of the tumors of origin, we assessed the effect of compounds **1**, **2**, and **3** on the colorectal carcinoma cell line HCT116 and the

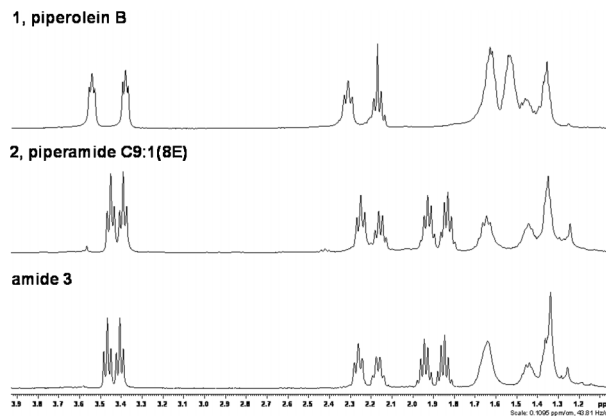


Fig. 2 ^1H NMR spectra showing the clear difference between piperidine and pyrrolidine amides.

urothelial (urinary bladder) carcinoma cell line EJ. We have previously reported that these cell lines are highly representative of the original tumors of origin and have characterized their responses to anti-proliferative (*pro*-apoptotic) responses *in vitro*.¹¹ This allowed us not only to determine the effect of the compounds on relevant representative cell lines but also permitted cell type-specific distinctions to be made. In addition to examining cytotoxic effects on cell cultures of standard cell density, *i.e.* $5\text{--}6 \times 10^3$ cells per well,¹² we also tested for biological effects using cultures of lower cell density (3×10^3 cells per well) in order to compare our findings to a previous report in which cytotoxicity detection assays were performed using relatively low density cultures of carcinoma (HeLa) cells to study the effects of isopiperolein B (Fig. 3).⁸

Piperolein B, **1** and piperamide C9:1(8E), **2** showed little (if any) detectable cytotoxicity at concentrations below $10 \mu\text{M}$. In comparison, amide **3** showed some cytotoxicity at $10 \mu\text{M}$, more significant effects at $30 \mu\text{M}$ and it caused complete cellular death in all cultures at higher concentrations ($100 \mu\text{M}$). This was independent of the initial seeding density of the cell cultures. All compounds demonstrated cytotoxicity at concentrations over $30 \mu\text{M}$ but only compound **3** resulted in 100% loss of viability in both cell lines and was hence the most cytotoxic independent of cell culture density and cell type. Interestingly, treatment with compound **1** at $100 \mu\text{M}$ caused 100% death only in HCT116 cells but not in EJ cells, indicating a differential effect between the cell lines. This however was only noticeable when cells were cultured at normal (but not low) density. Such a cell type-dependent differential response was even more pronounced in the case of compound **2**; at high doses, the compound showed almost 100% toxicity in HCT116 cells but the effect was much less pronounced in EJ cells which showed 60% cell viability.

Morphological observation of the cultures by routine, phase contrast microscopy indicated that compound **3** caused rapid and extensive cell death within less than 24 h after treatment and complete death in cultures of both cell lines by 48 h. The effect of the other two compounds was slower, and particularly in the case of compound **2** for EJ cells only a proportion of cells

Table 1 Comparison of NMR data reported for isopiperolein B and that obtained for amide **3**

^1H lit. data	^1H amide 3	^{13}C lit. data	^{13}C amide 3
1.35–1.50, 14H	1.29–1.50, 8H 1.56–1.71, 2H	24.5 25.4	24.8 25.3
	1.85, 2H, pent	26.5	26.5
	1.95, 2H, pent	28.9	29.4
2.15, 2H, q	2.12–2.21, 2H	29.3	29.7
2.30, 2H, t	2.26, 2H, t	29.7	29.8
3.40, 2H, t	3.41, 2H, t	32.8	33.3
3.55, 2H, t	3.47, 2H, t	33.4	35.2
5.90, 2H, s	5.93, 2H, s	42.5	45.9
5.95–6.15, 1H	6.04, 1H, dt	46.6	47.0
6.25, 1H, d	6.28, 1H, d	100.8	101.2
6.60–7.00, 3H	6.71–6.89, 3H	106.0	105.7
		108.5	108.5
		119.6	120.5
		128.0	129.6
		129.2	129.7
		132.4	132.8
		146.4	146.8
		147.8	148.2
		171.4	172.3



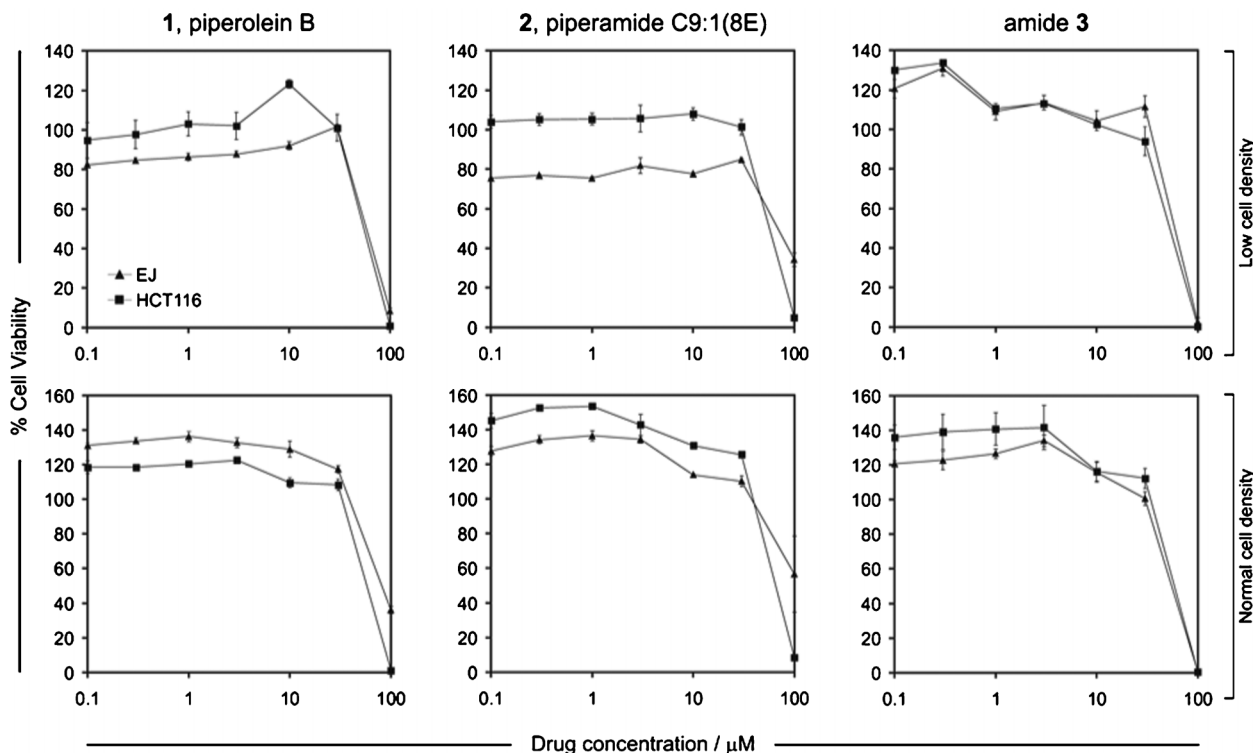


Fig. 3 The effect of the synthetic compounds **1**, **2** and **3** on carcinoma cell viability. HCT116 and EJ carcinoma cells were grown at 3×10^3 cells per well (low cell density) and 6×10^3 cells per well (normal cell density), before being treated with the indicated doses of compounds **1**, **2** and **3**. Cell viability (expressed as % in relation to controls) was assessed 48 h later as described in the experimental section and results are expressed as mean % values (\pm standard error of the mean). Results are mean values for six technical replicates and are representative of three independent experiments.

appeared fully apoptotic (dead) and a fraction of these remained alive.

Our findings indicate that the choice of initial cell density might be important if subtle cytotoxicity capacities and cell type-specific effects are to be revealed. The example of compound **1** exemplifies this point: the use of low cell density indicated complete cytotoxicity, yet the use of normal (standard) density revealed a dose and cell type-specific response. Thus the use of cultures that are of too low confluence might mask cell type specific responses and also suggest misleadingly high cytotoxic capacity.

It should also be noted that when treated at sub-lethal drug concentrations, most cultures showed an increased level of growth, *i.e.* over 100% cell viability in comparison to negative controls. This is not an anomalous observation that was specific to our experiments. Our recent *in vitro* findings (unpublished) and observations by others¹³ both *in vitro* and *in vivo* suggest that both normal and cancer epithelial cells exhibit a biphasic response to cytotoxic chemicals, consisting of increased growth at sub-cytotoxic drug concentrations, yet dramatic cytotoxicity at higher doses. This appears to be an inherent ability of epithelial cells to respond to such 'insults' as part of a repair response.^{13a}

Collectively, our results demonstrate for the first time that all of the three compounds exhibit differential effects on carcinoma cells and these effects appear to be both dose- and

cell type-dependent. Of the three compounds, **3**, the reported structure of isopiperolein B, was the most cytotoxic and piperamide C9:1(8E), **2** was the least cytotoxic demonstrating that the length of the connecting alkyl chain has an impact on activity. Consideration of the structures and the observed differences in their cytotoxic capacities could be used as a basis for structure-activity studies as well as for detailed biological investigations in order to understand the structural basis for these activities. In this way, the design of compounds with improved activities and pharmacological properties for anticancer therapy should be possible.

Conclusion

A simple synthetic strategy to three structurally related *Piper* compounds has been developed and their cytotoxic properties studied. Critically, the assigned structure of isopiperolein B has been shown not to be that of amide **3**, although compound **3** was the most cytotoxic compound of the three tested.

Experimental

¹H NMR spectra were recorded at 400 MHz. Chemical shifts are reported in ppm from tetramethylsilane with the solvent resonance as the internal standard (CDCl₃: 7.26 ppm). ¹³C



NMR were recorded with complete proton decoupling. Chemical shifts are reported in ppm from tetramethylsilane with the solvent as the internal standard (CDCl_3 : 77.4 ppm). Mass spectrometry (m/z) was performed in ESI mode, with only molecular ions being reported. Infrared (IR) spectra ν_{max} are reported in cm^{-1} . All purchased reagents were used as received without further purification. Petroleum ether refers to the fraction boiling at 40–60 °C. All reactions were performed under a N_2 atmosphere.

General procedure for amidation

1-Propanephosphonic acid anhydride solution (T3P; 50% in DMF, 1.39 mL, 4.8 mmol, 3 equiv.) was dissolved in CH_2Cl_2 (5 mL) and cooled to 0 °C. Triethylamine (1.3 mL, 9.6 mmol, 6 equiv.) and the carboxylic acid (1.6 mmol, 1 equiv.) were added. The reaction mixture was left to stir at 0 °C for 0.5 h, then piperidine or pyrrolidine (1.6 mmol, 1 equiv.) was added. After stirring overnight at room temperature the reaction was quenched with H_2O (20 mL) and extracted with EtOAc (3 × 10 mL). The combined organics were dried (MgSO_4), filtered and concentrated under vacuum to provide the corresponding amide. The crude products were deemed to be sufficiently pure and no further purification was performed.

1-(Piperidin-1-yl)non-8-en-1-one, 6. Isolated as a yellow oil. IR (neat): 1434 (s), 1639 (s), 2854 (m), 2926 (s) cm^{-1} . ^1H NMR (400 MHz, CDCl_3): δ 1.21–1.39 (6H, m), 1.42–1.64 (8H, m), 1.98 (2H, q, $J = 6.8$ Hz), 2.26 (2H, t, $J = 7.5$ Hz), 3.34 (2H, t, $J = 5.2$ Hz), 3.49 (2H, t, $J = 5.2$ Hz), 4.86 (1H, d, $J = 10$ Hz), 4.93 (1H, d, $J = 17$ Hz), 5.67–5.81 (1H, m). ^{13}C NMR (100 MHz, CDCl_3): δ 24.8, 25.7, 25.8, 26.8, 29.0, 29.1, 29.6, 33.7, 34.0, 42.9, 47.0, 114.4, 139.3, 171.9. MS: m/z ($M + 23$) 246.2 HRMS: m/z calc'd for $\text{C}_{14}\text{H}_{25}\text{NNaO}$ 246.1828, found 246.1822.

1-(Pyrrolidin-1-yl)non-8-en-1-one, 7. Isolated as a yellow oil. IR (neat): 1425 (s), 1637 (s), 2925 (m) cm^{-1} . ^1H NMR (400 MHz, CDCl_3): δ 1.22–1.38 (6H, m), 1.59 (2H, pent, $J = 7.1$ Hz), 1.79 (2H, pent, $J = 7.0$ Hz), 1.90 (2H, pent, $J = 6.4$ Hz), 1.98 (2H, q, $J = 7.0$ Hz), 2.20 (2H, t, $J = 7.5$ Hz), 3.36 (2H, t, $J = 6.8$ Hz), 3.41 (2H, t, $J = 6.9$ Hz), 4.82–4.98 (2H, m), 5.67–5.81 (1H, m). ^{13}C NMR (100 MHz, CDCl_3): δ 24.7, 25.1, 26.4, 29.0, 29.2, 29.6, 34.0, 35.0, 45.9, 46.9, 114.4, 139.3, 172.2. MS: m/z ($M + 23$) 232.2 HRMS: m/z calc'd for $\text{C}_{13}\text{H}_{23}\text{NNaO}$ 232.1672, found 232.1680.

1-(Pyrrolidin-1-yl)dec-9-en-1-one, 8. Isolated as a yellow oil. IR (neat): 1037 (m), 1442 (s), 1489 (s), 1636 (s), 2927 (m) cm^{-1} . ^1H NMR (400 MHz, CDCl_3): δ 1.22–1.41 (8H, m), 1.56–1.68 (2H, m), 1.83 (2H, q, $J = 6.8$ Hz), 1.93 (2H, pent, $J = 6.8$ Hz), 1.98–2.07 (2H, m), 2.23 (2H, t, $J = 7.8$ Hz), 3.39 (2H, t, $J = 6.8$ Hz), 3.44 (2H, t, $J = 6.8$ Hz), 4.91 (1H, d, $J = 10$ Hz), 4.97 (1H, d, $J = 17$ Hz), 5.72–6.86 (1H, m). ^{13}C NMR (100 MHz, CDCl_3): δ 24.7, 25.2, 26.4, 29.2, 29.3, 29.6, 29.8, 34.1, 35.2, 45.9, 46.9, 114.5, 139.5, 172.2. MS: m/z ($M + 23$) 246.2 HRMS: m/z calc'd for $\text{C}_{14}\text{H}_{25}\text{NNaO}$ 246.1828, found 246.1831.

General procedure for cross metathesis

Amide (0.20 mmol, 1 equiv.) and 5-vinylbenzo[*d*][1,3]dioxole **9** (120 mg, 0.81 mmol, 4 equiv.) were dissolved in 1,2-DCE (5 mL). Grubbs' second generation catalyst (18 mg, 0.020 mmol, 0.1 equiv.) was added and the mixture was heated at reflux for 2 days. The reaction mixture was concentrated under vacuum

and the residue purified by flash chromatography (2 : 1 petroleum ether–EtOAc).

Piperolein B, 1. Isolated as a yellow oil. IR (neat): 1037 (m), 1442 (s), 1489 (s), 1636 (s), 2927 (m) cm^{-1} . ^1H NMR (400 MHz, CDCl_3): δ 1.29–1.73 (14H, m), 2.17 (2H, q, $J = 7.0$ Hz), 2.31 (2H, t, $J = 7.5$ Hz), 3.38 (2H, t, $J = 5.4$ Hz), 3.54 (2H, t, $J = 5.4$ Hz), 5.93 (2H, s), 6.03 (1H, dt, $J = 16, 6.8$ Hz), 6.28 (1H, d, $J = 16$ Hz), 6.69–6.78 (2H, m), 6.89 (1H, s). ^{13}C NMR (100 MHz, CDCl_3): δ 25.0, 25.8, 26.0, 27.0, 29.4, 29.7, 29.8, 33.2, 33.9, 43.0, 47.1, 101.3, 105.8, 108.6, 120.6, 129.7 (2C), 132.8, 146.9, 148.3, 171.8. MS: m/z ($M + 23$) 366.2 HRMS: m/z calc'd for $\text{C}_{21}\text{H}_{29}\text{NNaO}_3$ 265.1563, found 366.2025.

Piperamide C9:1(8E), 2. Isolated as a yellow oil. ^1H NMR (400 MHz, CDCl_3): δ 1.28–1.51 (6H, m), 1.57–1.70 (2H, m), 1.83 (2H, pent, $J = 6.7$ Hz), 1.93 (2H, pent, $J = 6.5$ Hz), 2.16 (2H, q, $J = 7.0$ Hz), 2.25 (2H, t, $J = 7.0$ Hz), 3.39 (2H, t, $J = 6.7$ Hz), 3.45 (2H, t, $J = 6.8$ Hz), 5.92 (2H, s), 6.03 (1H, dt, $J = 16, 6.5$ Hz), 6.27 (1H, d, $J = 16$ Hz), 6.69–6.76 (2H, m), 6.88 (1H, s). ^{13}C NMR (100 MHz, CDCl_3): δ 24.8, 25.2, 26.5, 29.3, 29.6, 29.7, 33.2, 35.2, 46.0, 47.0, 101.2, 105.7, 108.5, 120.5, 129.6, 129.7, 132.8, 146.8, 148.2, 172.3. MS: m/z ($M + 23$) 352.2 HRMS: m/z calc'd for $\text{C}_{20}\text{H}_{27}\text{NaO}_3$ 352.1883, found 352.1871.

Proposed structure of isopiperolein B, 3. Isolated as a yellow oil. IR (neat): 1036 (s), 1248 (s), 1444 (s), 1605 (m), 1728 (m), 2926 (m) cm^{-1} . ^1H NMR (400 MHz, CDCl_3): δ 1.29–1.50 (8H, m), 1.56–1.71 (2H, m), 1.85 (2H, pent, $J = 7.0$ Hz), 1.95 (2H, pent, $J = 7.0$ Hz), 2.12–2.21 (2H, m), 2.26 (2H, t, $J = 7.9$ Hz), 3.41 (2H, t, $J = 6.5$ Hz), 3.47 (2H, t, $J = 7.0$ Hz), 5.93 (2H, s), 6.04 (1H, dt, $J = 16, 7.0$ Hz), 6.28 (1H, d, $J = 16$ Hz), 6.71–6.78 (2H, m), 6.89 (1H, s). ^{13}C NMR (100 MHz, CDCl_3): δ 24.8, 25.3, 26.5, 29.4, 29.7, 29.8 (2C), 33.3, 35.2, 45.9, 47.0, 101.2, 105.7, 108.5, 120.5, 129.6, 129.8, 132.8, 146.8, 148.2, 172.2. MS: m/z ($M + 23$) 366.2 HRMS: m/z calc'd for $\text{C}_{21}\text{H}_{29}\text{NNaO}_3$ 366.2040, found 366.2027.

Assessment of compound cytotoxicity

The colorectal carcinoma HCT116 and bladder carcinoma EJ cell lines were used to assess the effect of the synthetic compounds on cancer cell viability. Cells were seeded in 96-well plates at low (3×10^3 cells per well) and normal (6×10^3 cells per well) density in culture medium, which was a 1 : 1 (v/v) mixture of DMEM and RPMI (SigmaAldrich) supplemented with 5% (v/v) fetal bovine serum. Cells were incubated at 37 °C in a humidified atmosphere containing 5% (v/v) CO_2 and permitted to adhere overnight. Following synthesis, the compounds were aseptically reconstituted in DMSO at a final concentration of 20 mM. For testing, all compounds were diluted to concentrations 0.1, 0.3, 1, 3, 10 and 30 and 100 μM in culture medium. For each condition, six replicate samples were prepared. Solvent-alone treatments were also included for comparison (controls). Cells were then cultured for a period of 48 h before cell viability assays were performed. To assess cell viability, the CellTiter 96® AQueous One Solution Cell Proliferation Assay (Promega) was used, which is a colorimetric MTS viability assay based on the principle that viable cells have the ability to bioreduce the MTS tetrazolium compound into a coloured formazan product which is soluble in culture medium. Following drug treatment of cells for 48 h, CellTiter reagent was added to each well (as recommended by



the manufacturer). After a 4-hour incubation at 37 °C, cell viability was determined by measuring absorbance at 492 nm on a FLUOStar OPTIMA plate reader (BMG LabTech). Absorbance readings were obtained following background subtraction and % cell viability was calculated for each drug concentration in comparison to controls.

Notes and references

- For reviews, see: (a) J. C. do Nascimento, V. F. de Paula, J. M. David and J. P. David, *Quim. Nova*, 2012, **35**, 2288; (b) V. S. Parmar, S. C. Jain, K. S. Bisht, R. Jain, P. Taneja, A. Jha, O. D. Tyagi, A. K. Prasad, J. Wengel, C. E. Olsen and P. M. Boll, *Phytochemistry*, 1997, **46**, 597.
- F. Kiuchi, N. Nakamura, Y. Tsuda, K. Kondo and H. Yoshimura, *Chem. Pharm. Bull.*, 1988, **36**, 2452.
- (a) H. Matsuda, K. Ninomiya, T. Morikawa, D. Yasuda, I. Yamaguchi and M. Yoshikawa, *Bioorg. Med. Chem.*, 2009, **17**, 7313; (b) H. Matsuda, K. Ninomiya, T. Morikawa, D. Yasuda, I. Yamaguchi and M. Yoshikawa, *Bioorg. Med. Chem. Lett.*, 2008, **18**, 2038.
- (a) S. W. Lee, M.-C. Rho, H. R. Park, J.-H. Choi, J. Y. Kang, J. W. Lee, K. Kim, H. S. Lee and Y. K. Kim, *J. Agric. Food Chem.*, 2006, **54**, 9759; (b) S. Tsukamoto, K. Tomise, K. Miyakawa, B.-C. Cha, T. Abe, T. Hamada, H. Hirota and T. Ohta, *Bioorg. Med. Chem.*, 2002, **10**, 2981.
- Y. Okumura, M. Narukawa, Y. Iwasaki, A. Ishikawa, H. Matsuda, M. Yoshikawa and T. Watanabe, *Biosci., Biotechnol., Biochem.*, 2010, **74**, 1068.
- P. V. Srinivas and J. M. Rao, *Phytochemistry*, 1999, **52**, 957.
- S. V. Reddy, P. V. Srinivas, B. Praveen, K. H. Kishore, B. C. Raju, U. S. Murthy and J. M. Rao, *Phytochemistry*, 2004, **11**, 697.
- G.-H. Tang, D.-M. Chen, B.-Y. Qiu, L. Sheng, Y.-H. Wang, G.-W. Hu, F.-W. Zhao, L.-J. Ma, H. Wang, Q.-Q. Huang, J.-J. Xu, C.-L. Long and J. Li, *J. Nat. Prod.*, 2011, **74**, 45.
- (a) G. M. Strunz and H. J. Finlay, *Can. J. Chem.*, 1996, **74**, 419; (b) G. M. Strunz and H. J. Finlay, *Phytochemistry*, 1995, **39**, 731; (c) G. M. Strunz and H. J. Finlay, *Tetrahedron*, 1994, **50**, 11113.
- (a) K. Wei, S. Wang, Z. Liu, Y. Du, X. Shi, T. Qi and S. Ji, *Tetrahedron Lett.*, 2013, **54**, 2264; (b) M. Takahashi, M. Ichikawa, S. Aoyagi and C. Kibayashi, *Tetrahedron Lett.*, 2005, **46**, 57.
- (a) N. T. Georgopoulos, A. Merrick, N. Scott, P. J. Selby, A. Melcher and L. K. Trejdosiewicz, *Int. J. Cancer*, 2007, **121**, 1373; (b) U. Bugajska, N. T. Georgopoulos, J. Southgate, P. W. Johnson, P. Graber, J. Gordon, P. J. Selby and L. K. Trejdosiewicz, *J. Natl. Cancer Inst.*, 2002, **94**, 1381; (c) N. T. Georgopoulos, L. P. Steele, M. J. Thomson, P. J. Selby, J. Southgate and L. K. Trejdosiewicz, *Cell Death Differ.*, 2006, **13**, 1789; (d) L. P. Steele, N. T. Georgopoulos, J. Southgate, P. J. Selby and L. K. Trejdosiewicz, *Cell Death Differ.*, 2006, **13**, 1564.
- N. T. Georgopoulos, L. A. Kirkwood, D. C. Walker and J. Southgate, *PLoS One*, 2010, **5**, e13621.
- (a) R. Paus, I. S. Haslam, A. A. Sharov and V. A. Botchkarev, *Lancet Oncol.*, 2013, **14**, e50; (b) E. Bodo, D. J. Tobin, Y. Kamenisch, T. Biro, M. Berneburg, W. Funk and R. Paus, *Am. J. Pathol.*, 2007, **171**, 1153.

